



Ervilia castanea (Mollusca, Bivalvia) populations adversely affected at CO₂ seeps in the North Atlantic

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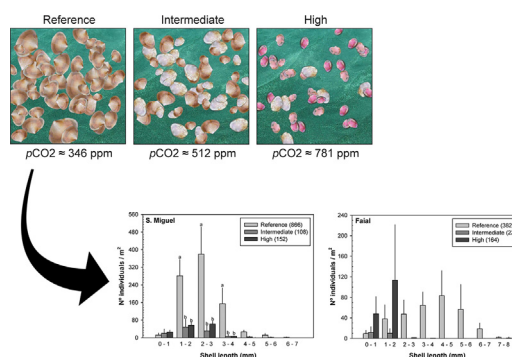
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HIGHLIGHTS

- The bivalve *Ervilia castanea* was studied at volcanic CO₂ seeps and reference sites.
- Abundance, size and net-calcification were inversely related to CO₂ levels.
- Large individuals were scarce or absent at high CO₂ sites.
- Recruitment of this bivalve was highest at the CO₂ seeps.
- Abundance and size of *E. castanea* were positively correlated with Chl-a in sediment.

GRAPHICAL ABSTRACT



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ABSTRACT

Sites with naturally high CO₂ conditions provide unique opportunities to forecast the vulnerability of coastal ecosystems to ocean acidification, by studying the biological responses and potential adaptations to this increased environmental variability. In this study, we investigated the bivalve *Ervilia castanea* in coastal sandy sediments at reference sites and at volcanic CO₂ seeps off the Azores, where the pH of bottom waters ranged from average oceanic levels of 8.2, along gradients, down to 6.81, in carbonated seawater at the seeps. The bivalve population structure changed markedly at the seeps. Large individuals became less abundant as seawater CO₂ levels rose and were completely absent from the most acidified sites. In contrast, small bivalves were most abundant at the CO₂ seeps. We propose that larvae can settle and initially live in high abundances under elevated CO₂ levels, but that

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high rates of post-settlement dispersal and/or mortality occur. *Ervilia castanea* were susceptible to elevated CO₂ levels and these effects were consistently associated with lower food supplies. This raises concerns about the effects of ocean acidification on the brood stock of this species and other bivalve molluscs with similar life history traits.

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1. Introduction

Increasing atmospheric CO₂ concentrations have caused the pH of the surface ocean to fall by 0.1 units since the pre-industrial age (Bindoff et al., 2019; Dupont and Pörtner, 2013; Feely et al., 2009; Keeling et al., 2017) and further decreases, between 0.1 (RCP2.6) or 0.3 (RCP8.5) pH units, are expected by 2100 (Bindoff et al., 2019). This process of ocean acidification affects calcification of marine organisms, leading to permanent damage and reduced viability (Shirayama, 2005) and causing major changes to marine ecosystems (Doney et al., 2012). Shallow-water CO₂ seeps provide analogues for the effects of chronic ocean acidification, at spatial and temporal scales that are difficult to mimic in aquaria or mesocosms (Hall-Spencer and Harvey, 2019). Care is needed to assess potential anomalies in temperature, alkalinity, metals and H₂S which are frequently found at these volcanic sites and can also influence biological responses, but are not associated with ocean acidification (Vizzini et al., 2013; Mishra et al., 2020).

This type of in situ approach has shown that ocean acidification can lead to ecosystem simplification and reorganization of marine communities (Fabricius et al., 2015; Kroeker et al., 2013a, 2011; Agostini et al., 2018). The structure of benthic marine communities changes greatly when they are exposed to carbonated seawater (Widdicombe et al., 2009), with decreases in biodiversity, biomass and trophic complexity (Hall-Spencer and Harvey, 2019). Communities tend to change from being dominated by calcareous organisms to non-calcareous ones (Baggini et al., 2014; Christen et al., 2013; Hall-Spencer et al., 2008), granting the latter a perceived advantage in a future acidified ocean (Kroeker et al., 2013b; Vihtakari et al., 2013).

Organisms with calcareous shells and skeletons, such as coralline algae, molluscs and echinoderms, are expected to be adversely affected by increasing CO₂ levels (e.g., reductions in abundance or local extinctions; Kroeker et al., 2011; Ricevuto et al., 2012) due to susceptibility of the calcification process to reduced carbonate availability (Bindoff et al., 2019). Garilli et al. (2015) found two gastropod species dwarfed as an adaptation to carbonated seawater, which allowed them to maintain calcification and partially repair shell dissolution. Other studies show that pH (or pCO₂) on their own (apart from the carbonate saturation state) can cause physiological effects that result in less energy available for mineralization (Cyronak et al., 2016; Palmer, 1992; Roleda et al., 2012; Waldbusser et al., 2015).

The early life stages of molluscs are often particularly sensitive to ocean acidification (Clements et al., 2016; Green et al., 2013, 2004; Jansson et al., 2015; Ricevuto et al., 2012). Previous studies have shown stunted growth and shell development (Amaral et al., 2011; Gazeau et al., 2010; Hettinger et al., 2012; Kroeker et al., 2010; Talmage and Gobler, 2010; Waldbusser et al., 2015); reduced sperm motility, fertilization and hatching rates (Barros et al., 2013; Gazeau et al., 2010; Kroeker et al., 2010); slower metamorphosis and delayed settling (Jansson et al., 2015; Talmage and Gobler, 2010); decreased survival (Gazeau et al., 2010; Talmage and Gobler, 2010); changes to biochemical composition (Range et al., 2014a; Talmage and Gobler, 2010); increased frequency of shell abnormalities and dissolution (Barros et al., 2013; Green et al., 2004; Pereira et al., 2015; Talmage and Gobler, 2010); altered metabolic rates (Fernández-Reiriz et al., 2012, 2011; Mellado et al., 2019); altered burrowing behaviour and post-settlement dispersal (Clements et al., 2016) and; decreased settlement success (Gazeau et al., 2010). Furthermore, some species

reallocate energy budgets to cope with ocean acidification, with growth and reproduction often compromised to sustain fitness and survival (Clements et al., 2020; Durland et al., 2019; Widdicombe and Spicer, 2008). This type of trade-off has been found to produce negative (Dupont et al., 2012; Hettinger et al., 2012), positive (Parker et al., 2012; Zhao et al., 2018, 2019) and neutral (Clements et al., 2020) carry-over effects from the parental brood stocks to the next generation. These results indicate that the responses of the early life stages of bivalves to increased CO₂ go well beyond the detrimental effects on shell calcification.

Responses to ocean acidification are species-specific and even closely related species can react differently, depending on various factors, such as life history traits, habitat, sex, nutritional status, adaptation potential and methodological conditions (Calosi et al., 2013; Doney et al., 2012; Dupont and Thorndyke, 2009; Garilli et al., 2015; Kroeker et al., 2013a, 2011; Marčeta et al., 2020; Pörtner, 2008; Vihtakari et al., 2013). Individual responses to ocean acidification are of key importance for any given species, but they also have cascading effects along trophic relations and ecological interactions, making them important drivers in shaping community structures and phase shifts (Kroeker et al., 2011; Range et al., 2014b). Consequently, the direct effects of CO₂ on competitors, prey, predators and the quantity and quality of food supply can be translated to density control mechanisms (Metaxas, 2015; Range et al., 2012) and mediated by adaptation or acclimation (Kroeker et al., 2011).

The species targeted in this study, *Ervilia castanea* (Montagu, 1803), is a semelid bivalve (Morton, 1990) found from south of the British Isles into the Mediterranean (Babío and Bonnin, 1987). It has a brownish, oval and elongated shell, which reaches a maximum length of 15 mm (Macedo et al., 1999; Morton, 1990). In the Azores, where the majority of the bivalve species tend to be ~50% smaller than their continental conspecifics (Morton et al., 2013), the maximum length reported is 6.7 mm (Morton, 1990). This generalized dwarfism of bivalves is commonly found around oceanic islands and has been related to the low availability of food in these oligotrophic waters (Morton et al., 2013). Individuals of this species have a high potential reproductive output, with maturation occurring at a length of 3.5 mm for females and 5.5 mm for males (Morton, 1990). *Ervilia castanea* occupies subtidal coarse-grained, well-sorted sediments overlain by clean, nutrient poor, oceanic waters, to depths exceeding 40 m (Morton, 1990). In the Azores it is numerically dominant on sandy substrata and mainly predated upon by naticid gastropods (Morton, 1990; Morton et al., 2013).

In this study, we collected in situ evidence about the effects of elevated seawater CO₂ on populations of the bivalve *Ervilia castanea*, along submarine volcanic CO₂ gradients in the Azores. We recorded abundance, size-structure, biomass and tissue composition. Three hypotheses were tested: we expected the abundance (Hyp. 1) and net-calcification (i.e., accretion minus dissolution, Hyp. 2) of *Ervilia castanea* to decrease with rising CO₂ levels and for the size structure of the population (Hyp. 3) to be modified along the CO₂ gradients.

2. Methods

2.1. Selection of the study sites

The Azores are nine volcanic islands located on the triple junction of the Eurasian, North-American and Nubian tectonic plates. Shallow-water volcanic CO₂ seeps occur around several of the Azorean islands

and seamounts (Couto et al., 2015; Rajasabapathy et al., 2014; Tribollet et al., 2018; Viveiros et al., 2016; Wallenstein et al., 2013, 2009). Two of these coastal sites, located off the islands of São Miguel and Faial, were chosen for this study, based on their shallow depth and accessibility, as well as previous knowledge about the intensity and composition of the gas emissions. Ponta da Lobeira (São Miguel, 37°43'31.8"N 25°19'01.6"W, Supplementary Video 1) is a site with strong degassing at 7 m depth, located <2 km from the village of Ribeira Quente, on the south flank of Furnas Volcano. Ponta da Espalamaca (Faial, 38°32'36.4"N 28°35'48.4"W, Supplementary Video 2) is a site with more diffuse degassing, at 37 m depth, located 3 km from Horta port (Fig. 1). Supplementary videos one and two show the type of benthic habitat and the degassing fluxes at Ponta da Lobeira and Ponta da Espalamaca, respectively.

2.2. Experimental design

Off each island, three sampling sites, with similar depth, bottom topography exposure to currents and wave action, were selected: a significant degassing site (High CO₂); a transitional site (Intermediate CO₂), where the seawater carbonate chemistry was still affected, but with no visible CO₂ release; and a reference site (Reference), unaffected by the CO₂ emissions. Test dives with a pH multiprobe (YSI6600) were used to define these sites. In São Miguel, sampling was done between 3 and 7 July 2014, at depths ranging between 6 and 10 m. All sites in São Miguel were relatively sheltered from the dominant wave action. Sampling in Faial was done between 10 and 15 July 2014, at depths between 28 and 37 m. Given the proximity to the Faial-Pico channel, all the sites in Faial were strongly exposed to tidal currents.

2.3. Sample collection and laboratory procedures

2.3.1. Free gas composition and flux

Free gas samples were collected at the High CO₂ sites using Giggenbach bottles (Supplementary Video 3) and gas composition was analysed by titration and gas chromatography (for additional details, Ferreira et al., 2005). Inverted plastic funnels, connected to volumetric flasks, were used to measure gas emission rates. In São Miguel (Ponta da Lobeira), this sampling was stratified into low, medium and high flux areas, defined by visual inspection of bubbling, with three replicate samples collected at each level. In Faial (Ponta da Espalamaca), intensity of bubbling was more homogeneous, so the different levels of emission could not be established and two replicates were collected in each of three haphazardly selected points of emission (A, B and C).

2.3.2. Physicochemical properties of seawater

Seawater temperature, salinity, dissolved oxygen and total pH (pH_t) were measured at least once per day at each sampling site, over the course of 3 or 4 days. A YSI6000 multiprobe was used for continuous (5 second intervals) in situ measurements during vertical profiles (3 days per site) and diving operations (1 or 2 days per site). A Niskin bottle was also used during the vertical profiles, for collecting discrete samples of sub-surface (0–2 m) and near-bottom (6–10 m in São Miguel and 15–38 m in Faial) seawater, for the determinations of dissolved inorganic nutrients, total pH (pH_t) and total alkalinity (TA). pH_t was measured from samples without headspace using a glass electrode (WTW 340i pH meter) and calibrated with a Tris seawater buffer provided by the A. Dickson laboratory. TA samples were filtered through a 0.2 µm pore size, stored at 4 °C and measured within 48 h by potentiometric titration according to Dickson et al. (2003) using a Metrohm 848 Titrino Plus equipped with Metrohm 869 Compact Changer Sampler. TA measurements were corrected with certified reference material (Dickson, 2010), at 20 µmol·kg⁻¹ accuracy and 2 µmol·kg⁻¹ precision. Samples for the determination of dissolved inorganic silicate and phosphate were filtered using a 0.2 µm polyethersulfone syringe filter and stored at -20 °C until analysis. Concentrations of silicate and phosphate

were measured following Hansen and Koroleff (1999), by means of a spectrophotometer (Cary 60, Varian). Seawater carbonate chemistry speciation was calculated from temperature, salinity, silicate, phosphate, multiprobe pH_t values and TA using CO₂SYS (Lewis and Wallace, 1998) with the equilibrium constants determined by Mehrbach et al. (1973) as refitted by Dickson and Millero (1987), by interpolating discrete variables from Niskin sampling (silicate, phosphate and TA) with continuous multiprobe records (pH_t, temperature and salinity). Accordingly, the calculations were done using the average values of TA, silicate and phosphate for each vertical profile (i.e., average of bottom and surface samples per sampling site and per day).

2.3.3. Macrofauna

Sediment macrofauna samples were collected by scientific divers, from an area of 0.25 m² excavated to 15 cm depth, using an airlift fitted with a 500 µm mesh bag (Supplementary Video 4). Replicate samples were spaced by 2 to 6 m and haphazardly positioned within each site, except in the High CO₂ sites, where the points of gas emission were explicitly avoided. In São Miguel, nine replicates were collected at each site. In Faial, time restrictions and operational limitations while diving at greater depths and in stronger currents, enabled the collection of only six samples of macrofauna in the Reference and Intermediate sites and seven in the degassing site.

All organisms retained in the bag were preserved in 5% buffered formaldehyde, hand-sorted and later preserved in 70% ethanol until taxonomic determination and counting. All the *Ervilia castanea* were individually measured and subsequently pooled per replicate and shell length (1 mm size-classes, 0 to 8 mm). These pools of bivalves were dried for a minimum of 36 h at 60 °C (dry weight - DW) and subsequently burnt in a muffle furnace for 4 h at 550 °C (ash weight - AW). Ash-free dry weight (AFDW) and relative ash weight (%AW) were estimated, per replicate and size-class using Eqs. (1) and (2), respectively:

$$\text{AFDW} = \frac{\text{Dry Weight} - \text{Ash Weight}}{\text{Dry Weight}} \quad (1)$$

$$\% \text{AW} = \frac{\text{Ash Weight}}{\text{Dry Weight}} \times 100 \quad (2)$$

2.3.4. Sediments

Hand-held PVC cores (3.5 cm diameter per 15 cm length) were used to collect two sediment samples, adjacent to each macrofauna sample. The first core was used for determination of granulometry and organic matter/carbonate content. A set of six sieves (63 µm, 125 µm, 250 µm, 500 µm, 1 mm and 2 mm) was used for sediment pooling per grain size. After sieving, each fraction was weighed and related to the total weight of the sample, as a percentage. The organic matter content and carbonate content in sediments was determined by the loss-of-ignition (LOI) method (Heiri et al., 2001). Sediment was dried in an oven for 72 h at 100 °C (DW₁₀₀), from which, 10 g of each sample were burnt for 8 h at 550 °C (DW₅₅₀) and subsequently for 2 h at 900 °C (DW₉₀₀). Organic matter content (LOI₅₅₀) and carbonate content (LOI₉₀₀) were then determined using Eqs. (3) and (4), respectively.

$$\text{LOI}_{550} = \frac{\text{DW}_{100} - \text{DW}_{550}}{\text{DW}_{100}} \times 100 \quad (3)$$

$$\text{LOI}_{900} = \frac{\text{DW}_{550} - \text{DW}_{900}}{\text{DW}_{100}} \times 100 \quad (4)$$

The second core was used for carbon/nitrogen ratio and photosynthetic pigment analyses. Sediment samples were frozen and kept in the dark for preservation of chloroplastic pigments. Approximately 12 g of frozen sediment were lyophilized in a Telstar LyoAlfa 15 for 48 h, to get the 5 g of material needed. For total organic carbon (TOC) and total nitrogen (TN) samples were homogenized and acidified with

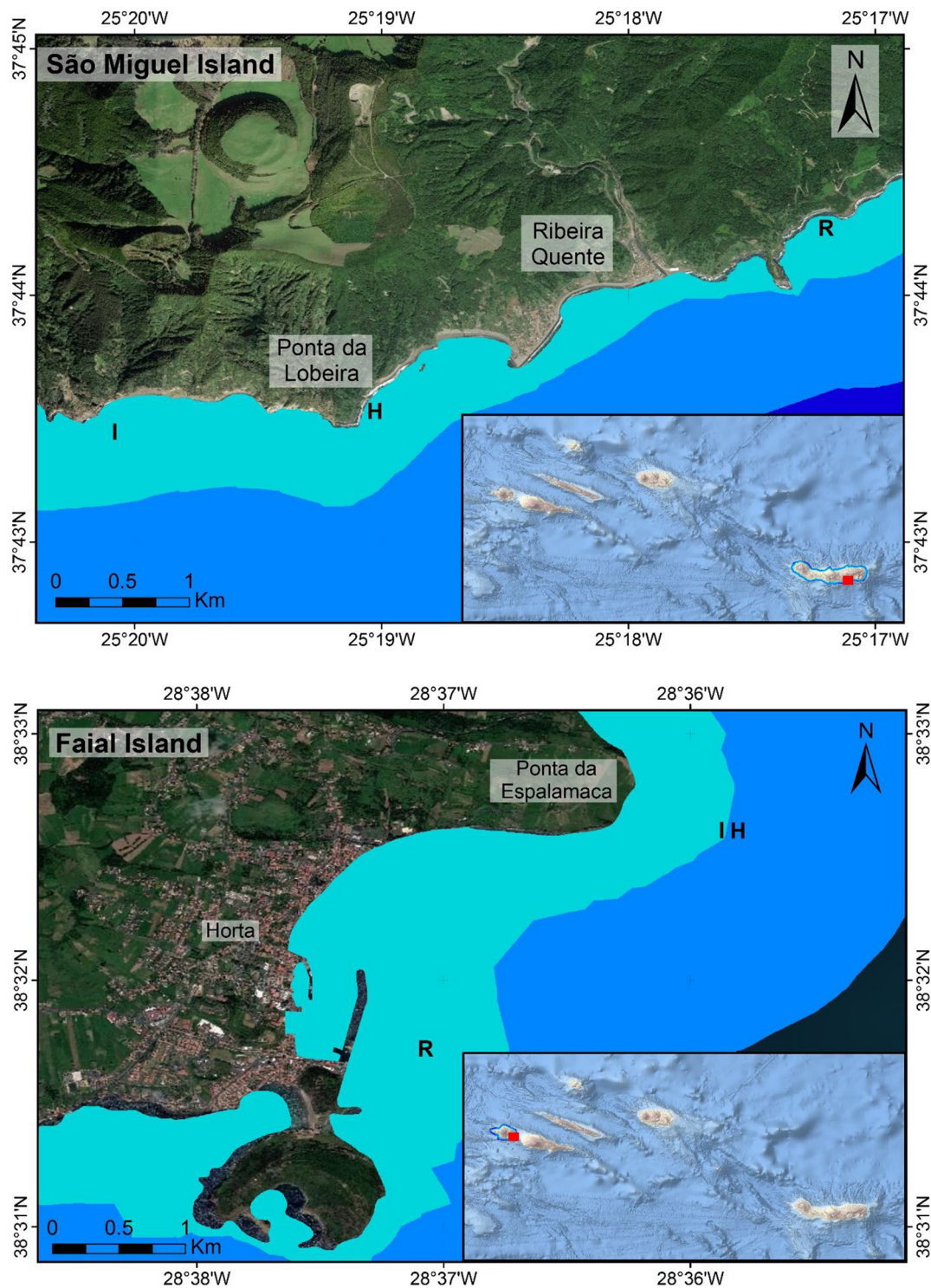


Fig. 1. Map of sampling locations in the central and eastern groups of the Azores archipelago: R – Reference, I – Intermediate CO₂, H – High CO₂. The blue gradient represents the 3 depth categories defined for coastal waters around the islands: 0–30 m, 30–200 m and > 200 m. (For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article.)

dilute hydrochloric acid until complete decarbonization. TOC was measured with a Thermo Scientific Flash 2000 elemental analyser. For analyses of the chloroplastic pigments, samples were homogenized and pigments extracted in 90% acetone. Pigment separation was done using reverse phase high performance liquid chromatography (HPLC) and measured with a Gilson fluorescence detector according to Wright and Jeffrey (1997).

2.4. Statistical analysis

Two-way analyses of variance (ANOVA) were used to test for differences in abundance and biomass of *Ervilia castanea* within each island (São Miguel and Faial). CO₂ level (High, Intermediate and Reference) and size-class were considered as fixed orthogonal factors. The data was untransformed and Cochran's test was used to test for

heterogeneity of variance. When appropriate, multiple comparisons for means were done on significant main effects using Student–Newman–Keuls (SNK) tests, with adjusted p -values for each pairwise comparison. Univariate analysis and graphs were done using SigmaPlot software (Version 13.0, Systat Software, Inc.) and R software (Version 3.1.1).

Distance-based linear modelling (DistLM) was used to investigate the relationship between sediment variables and the size structure of *Ervilia castanea* (Bray–Curtis similarity matrix). The significance of the relationship was first evaluated for individual variables with marginal tests (999 permutations). Significant variables were included in the model selection using BEST routine (i.e. all possible variable combinations). Distance-based redundancy analysis (dbRDA) was used for the ordination and visualization of the best overall DistLM solution, according to Akaike's Information Criteria (AIC). All the multivariate analyses were done using PRIMER 6 (Version 6.1.13) statistical package with the PERMANOVA add-on (Version 1.0.3).

3. Results

3.1. Free gas composition and flux

Free gas composition at both vent sites was mainly carbon dioxide. In São Miguel, CO₂ accounted for >99 mol% of gas composition, while in Faial CO₂ accounted for between 98 and 99 mol%. Nitrogen, argon and oxygen followed with values ranging between 0.02 mol% and 1.27 mol%. The remaining gases (helium, hydrogen sulfide, hydrogen and methane) had very low concentrations, with some, mainly in Faial, being absent or below detectable limits (Table 1).

3.2. Physicochemical properties of seawater

Seawater temperature and salinity varied between the two islands and with depth, but not among sampling sites within each island. Average salinity was higher in Faial (37.69 ± 0.05), relative to São Miguel (36.83 ± 0.05). Average sea surface temperature did not vary between the two Islands at the time of sampling (20.61 ± 0.14), but the bottom layer was slightly warmer in São Miguel (20.08 ± 0.20), relative to Faial (18.90 ± 0.19). The carbonate chemistry variables showed a similar trend in both islands, consistent with the predefined gradients. In São Miguel, pH_T ranged from 6.92 (High CO₂ site) to 8.16 (Reference site). In Faial pH_T ranged between 6.81 (High CO₂ site) and 8.2 (Reference site). The lowest pH_T values were always found at the degassing sites, while the highest values were similar across all the sampling sites (Fig. 2). pCO₂ and HCO₃[−] increased along the gradients, while CO₃, OH[−] and CaCO₃ saturation (calcite and aragonite) showed the inverse pattern. Undersaturation ($\Omega < 1$) of aragonite and calcite was observed for the degassing sites in both islands and for the intermediate site in Faial. These values were, however, restricted to the extremes of the frequency distribution (Fig. 2 and Supp. Figs. 1 and 2) and did not affect the central tendency metrics (Table 2). Undersaturated conditions were, therefore, relatively infrequent, even at the high CO₂ sites.

3.3. Population structure of *Ervilia castanea*

A total of 3134 *Ervilia castanea* individuals were collected during this study, representing 58% of the overall abundance of macrofauna. *E. castanea* was the dominant organism in all sampling sites, except for the Intermediate site in Faial, where Nematodes of the genus *Metoncholaimus* were numerically dominant. Densities of *E. castanea* up to $1140 \text{ ind} \cdot \text{m}^{-2}$ were recorded in São Miguel, which generally had larger abundances, relative to Faial (Fig. 3). In Faial, the size-frequency distribution was clearly skewed towards smaller individuals at the High CO₂ site, while in São Miguel the sites tended to have a more similar distribution. Average abundances (across all size classes) consistently peaked at reference sites and this effect was significant in both Islands (Table 3). In contrast, the smallest size class (<1 mm), composed of recently settled postlarvae (i.e., *plantigrades*, sensu Carriker, 1961) and juveniles (spat), showed consistently larger abundances in sites affected by CO₂ than in reference sites, which explains the significant interaction term for São Miguel. The dominant size class in Faial (1–2 mm) also showed a similar pattern, reaching peak abundance in the high CO₂ site (Fig. 3).

3.4. Biomass of *E. castanea*

The smallest (<2 mm) and largest (>5 mm) size classes were excluded from the biomass analyses, due to lack of precision in weighing and absence in the high CO₂ sites, respectively. The average biomass per individual (DW, AW and AFDW) increased with shell length for the three intermediate size classes (between 2 and 5 mm shell length, Fig. 4). This pattern was visible in all sampling sites, except for the intermediate site in Faial, where individuals larger than 2 mm were absent. Relative ash weight differed significantly among sites (%AW, Table 3), with the dominant size classes showing consistently larger values in reference sites (92% in both islands), than in sites affected by CO₂ emissions (83% in São Miguel and 75% in Faial). Although no quantification of shell dissolution or strength was done in this study, alterations were clearly apparent on the few larger individuals found at Intermediate and High CO₂ areas, particularly around the umbo, where it was usually associated with some degree of erosion to the periostracum (Fig. 5B and D).

3.5. Relationship between sediment characteristics and population structure of *E. castanea*

DistLM marginal tests indicated that 13 out of the 15 sediment variables considered were significantly related to the distribution and size structure of *E. castanea* (Table 4). The only exceptions were silt (sediment finer than 63 µm) and total nitrogen (TN). These two variables were, therefore, excluded from the model selection. The best overall DistLM solution included 4 sediment variables: 2 mm grain size, carbonate content, chlorophyll *a* and other pigments (diadinoxanthin, diatoxanthin, zeaxanthin and lutein). The coarser sediments were strongly associated with the intermediate and high CO₂ sites in Faial (Fig. 6), which also had the smallest abundances of *E. castanea* above 2 mm shell length (Fig. 3). Chlorophyll *a* was the dominant photosynthetic pigment in all sampling sites,

Table 1

Gas composition (mean values) of samples collected at each of the two degassing systems (Ponta da Lobeira – São Miguel, $n = 3$; Ponta da Espalamaca, Faial, $n = 2$). Carbon dioxide (CO₂), hydrogen sulfide (H₂S), methane (CH₄), hydrogen (H₂), helium (He), nitrogen (N₂), oxygen (O₂), argon (Ar). B.d.l. (below detection limit).

Site	Emission rate	CO ₂ (mol%)	H ₂ S (mol%)	CH ₄ (mol%)	H ₂ (mol%)	He (mol%)	N ₂ (mol%)	O ₂ + Ar (mol%)	Gas flux (L/min/m ²)
S. Miguel	Low	99.49	b.d.l.	1.14E^{-02}	1.39E^{-04}	1.01E^{-03}	0.37	0.13	0.58
S. Miguel	Medium	99.86	9.66E^{-04}	1.25E^{-02}	4.00E^{-05}	9.95E^{-04}	0.11	0.02	10.26
S. Miguel	High	99.58	8.97E^{-02}	1.39E^{-02}	3.97E^{-02}	1.06E^{-03}	0.23	0.05	24.88
Faial	A	98.61	b.d.l.	b.d.l.	b.d.l.	6.32E^{-03}	1.27	0.12	2.64
Faial	B	98.91	b.d.l.	b.d.l.	b.d.l.	3.41E^{-03}	0.94	0.15	2.64
Faial	C	99.35	b.d.l.	b.d.l.	b.d.l.	2.24E^{-03}	0.56	0.09	2.64

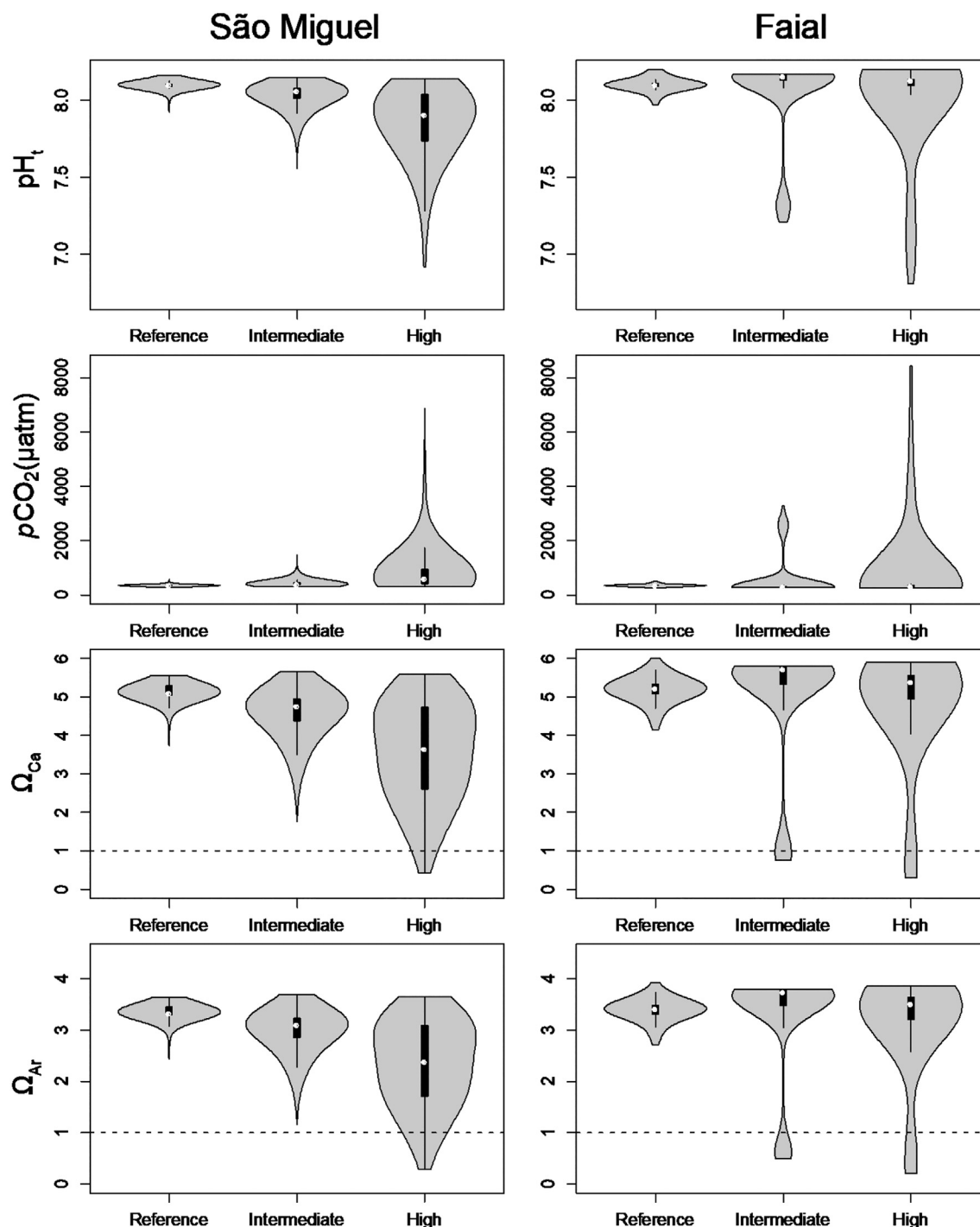


Fig. 2. Violin plots for pH (total scale), CO₂ partial pressure (pCO₂), saturation of calcite (Ω_{Ca}) and aragonite (Ω_{Ar}); the frequency distribution (grey area), median (white dot) and quartiles (black bars) are presented for each site in each island, across all depths and dates of sampling.

followed by fucoxanthin. All pigments showed higher concentrations in the reference sites, relative to the ones affected by CO₂, and this pattern was strongly associated with the intermediate size classes (2–5 mm shell length) of *E. castanea*. The reference site in Faial was also characterized by elevated values of carbonates (CaCO₃) and organic matter (OM), relative to all the other sampling sites (Table 4).

4. Discussion

Gas released from both seep systems was mainly CO₂ (>98.6%), with N₂, O₂, Ar and He also present in the dry phase. Residual fractions of

other typical hydrothermal gases (H₂S, H₂ and CH₄) were also detected at Ponta da Lobeira (Table 1). The sulfuric component was, however, only present when the gas flux was medium to high (≥10 L/min/m²) and in relatively small concentrations (<10 ppm), when compared to the sub-aerial fumaroles of the Furnas Volcano (Caliro et al., 2015) or to other submarine vents (Tarasov et al., 2005). Boatta et al. (2013) recorded much higher concentrations of H₂S (400 ppm) in the vent bubbles at Vulcano Island, Italy, concluding that only a small proportion of sulfide enters or remains in the aquatic phase and that the dissolved levels dropped below the detection limit at 5 m distance from the emission point. Accordingly, although we cannot dismiss the possibility of

Table 2

Seawater carbonate chemistry variables (mean \pm SE) at each sampling site (R – Reference, I – Intermediate CO₂, H – High CO₂) in São Miguel (SM) and Faial (FA). The variables represented are: temperature (Temp), salinity (Sal), pH (total scale), total alkalinity (TA), CO₂ (total scale), CO₂ partial pressure, bicarbonate (HCO₃⁻), carbonate (CO₃²⁻), ion hydroxide (OH⁻), saturation of calcite (Ω_{Ca}) and aragonite (Ω_{Ar}). Each entry represents the average of bottom and surface measurements per sampling site and per day (n = 3).

Location	Temp (°C)	Salinity (ppt)	pH _t	TA (μmol/kg)	TCO ₂ (μmol/kg)	pCO ₂ (μatm)	HCO ₃ ⁻ (μmol/kg)	CO ₃ ²⁻ (μmol/kg)	OH ⁻ (μmol/kg)	Ω_{Ca}	Ω_{Ar}
SM-R	20.13 \pm 0.01	36.93 \pm 0.00	8.10 \pm 0.00	2371.47 \pm 0.07	2056.80 \pm 0.27	347.80 \pm 0.45	1827.08 \pm 0.43	218.65 \pm 0.18	5.04 \pm 0.01	5.15 \pm 0.00	3.36 \pm 0.00
SM-I	20.06 \pm 0.01	36.98 \pm 0.00	8.03 \pm 0.00	2380.92 \pm 0.00	2104.04 \pm 1.02	430.63 \pm 2.69	1895.06 \pm 195.25	195.25 \pm 0.63	4.34 \pm 0.02	4.60 \pm 0.01	3.00 \pm 0.01
SM-H	20.54 \pm 0.00	36.94 \pm 0.00	7.86 \pm 0.00	2397.89 \pm 0.06	2199.16 \pm 1.42	818.95 \pm 10.07	2021.84 \pm 1.92	151.51 \pm 0.77	3.31 \pm 0.02	3.57 \pm 0.02	2.33 \pm 0.01
FA-R	20.14 \pm 0.04	37.65 \pm 0.01	8.10 \pm 0.00	2371.70 \pm 0.00	2050–46 \pm 0.85	345.94 \pm 1.59	1817.50 \pm 1.33	222.00 \pm 0.53	5.12 \pm 0.02	5.20 \pm 0.01	3.39 \pm 0.01
FA-I	19.91 \pm 0.05	37.62 \pm 0.01	8.04 \pm 0.01	2345.20 \pm 0.00	2044.58 \pm 6.31	594.01 \pm 38.00	1811.93 \pm 8.37	213.05 \pm 3.34	5.11 \pm 0.01	4.99 \pm 0.08	3.26 \pm 0.05
FA-H	19.58 \pm 0.05	37.56 \pm 0.01	8.02 \pm 0.01	2353.48 \pm 0.08	2069.69 \pm 4.52	744.14 \pm 44.22	1839.47 \pm 5.16	205.37 \pm 2.06	4.75 \pm 0.05	4.81 \pm 0.05	3.14 \pm 0.03

toxic gases, other than CO₂, affecting the bivalves at Ponta da Lobeira, this potential confounding factor was clearly less important here than in most previously studied shallow vent sites. Other types of contaminants usually present in vent emissions, such as trace metals, were not assessed in our study. Their adverse biological effects are, however, usually localized to a few hundred meters from the vents (Vizzini et al., 2013), so it is unlikely that they could have affected the Intermediate CO₂ sites.

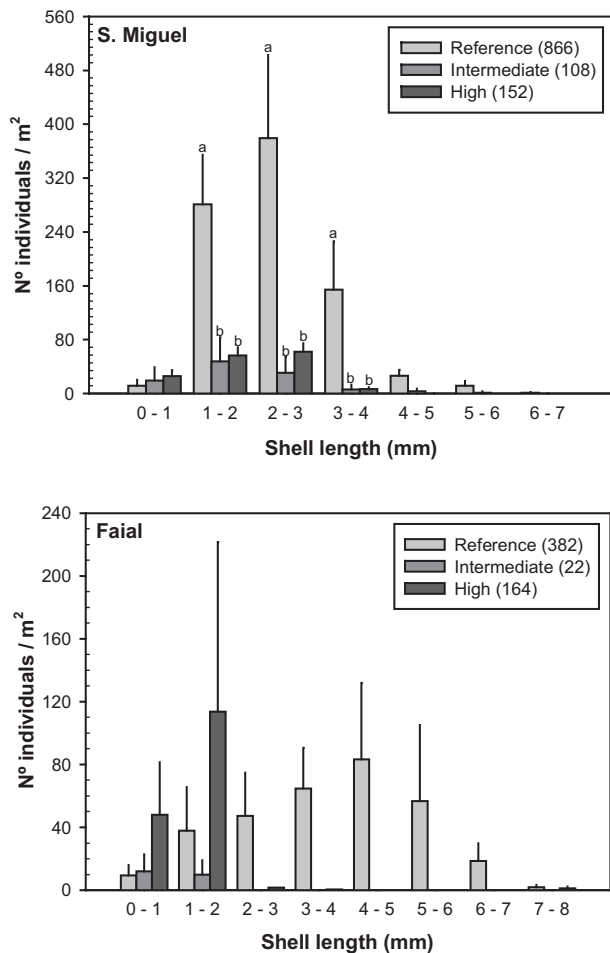


Fig. 3. Abundance (mean \pm SE) of the bivalve *Ervilia castanea* per size-class and CO₂ level in each of the islands; bars with different letters denote significant differences among CO₂ levels within each size class (SNK tests, $p < 0.05$); values in parenthesis represent mean abundances, across all size classes, at each CO₂ level in each of the islands.

Seawater temperature, salinity and total alkalinity varied within narrow ranges among the different sampling sites and these variations were independent of the CO₂ emissions. Seawater pH, on the other hand, varied substantially among sites. This was mainly due to the increased frequency of extremely low pH values (down to 6.8) at the sites affected by CO₂ emissions, rather than a consistent decrease of the average pH at those sites. Despite this, the gradient of CO₂ partial pressure in São Miguel was clearly within the range predicted by the IPCC for the end of this century, which varies between 421 (RCP2.6) and 936 (RCP8.5) ppm (Hoegh-Guldberg et al., 2014; Pörtner et al., 2014). The gradients in seawater carbonate chemistry observed in São Miguel can, therefore, be unambiguously applied to infer on the response of benthic organisms to present conditions (Reference site) and future scenarios of ocean acidification (Intermediate and High CO₂ sites). The gradients in carbonate chemistry were less clear for Faial, due to the greater depth, stronger currents and the fact that pH data was collected along the entire water column (37 m). These conditions complicate the interpretation, as variations exceed the average scenarios for ocean acidification in the foreseeable future. Nevertheless, the regulation of carbonate chemistry in coastal waters involves a multitude of drivers (i.e., watershed processes, nutrient inputs, changes in ecosystem structure and metabolism), apart from anthropogenic CO₂ emissions, so variations of this magnitude are not unusual for this type of habitat (Duarte et al., 2013). In fact, the potential adaptations of coastal ecosystems to this increased environmental variability, induced by interactions between global and local drivers, are critically important to

Table 3

Analyses of variance for the abundance and relative ash weight (%AW) of *Ervilia castanea* for each island; CO₂ level and Size Class are fixed orthogonal factors; df: degrees of freedom; MS: mean squares; # denotes groups of samples with heterogeneous variances (Cochran's tests, $p < 0.05$).

Abundance								
Source of variation	São Miguel [#]				Faial			
	df	MS	F-ratio	p	df	MS	F-ratio	p
CO ₂ level	2	14,529	20.166	<0.001	2	1042	3.342	0.038
Size Class	6	6945	9.64	<0.001	7	302	0.97	0.456
CO ₂ × Size	12	3408	4.73	<0.001	14	356	1.143	0.328
Residual	168	720			128	312		
Relative ash weight (%AW)								
Source of variation	São Miguel				Faial [#]			
	df	MS	F-ratio	p	df	MS	F-ratio	p
CO ₂ level	2	0.0697	5.009	0.01	1	0.0807	7.728	0.016
Size Class	2	0.0195	1.4	0.256	2	0.0294	2.814	0.097
CO ₂ × Size	4	0.0245	1.763	0.151	13	0.0104		
Residual	51	0.0139			16	0.0214		

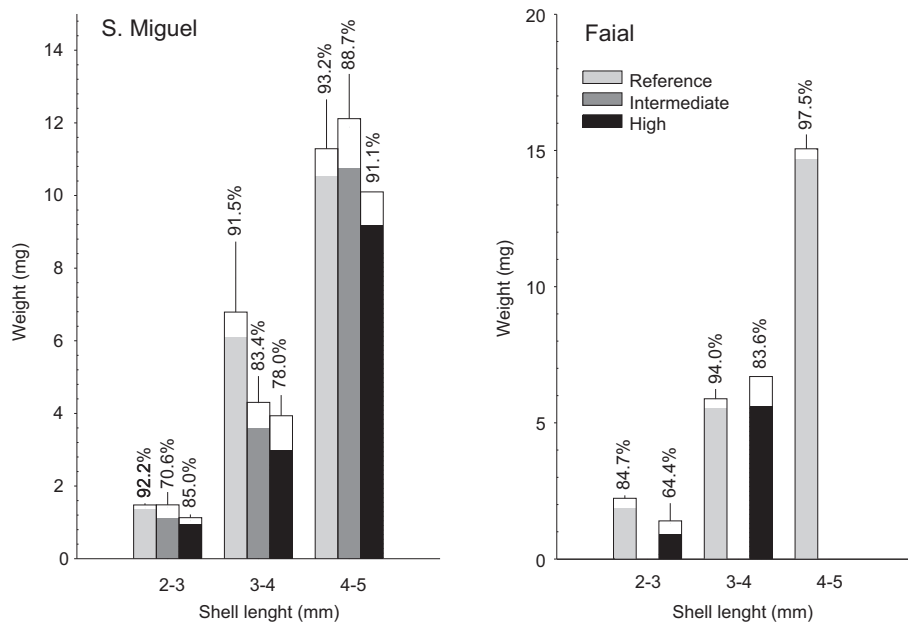


Fig. 4. Individual body weight of the dominant size classes of *Ervilia castanea* per CO₂ level in each Island; dry weight (outline, means \pm SE) and ash weight (filled area, mean); percentage values on top of the bars denote relative ash weight (mean).

forecast their sensitivity and vulnerability to future ocean acidification and cannot be ignored.

Hyp. 1 and Hyp. 2 were corroborated: *Ervilia castanea* significantly decreased in abundance and net-calcification, measured as relative ash weight, with increasing CO₂ levels. Individuals of all sizes were represented in reference sites, while larger individuals (above 3 mm shell length) were scarce or absent in sites affected by CO₂ emissions. This steep decline in the abundance of larger individuals was probably caused by high levels of post-settlement mortality, as a consequence of delayed or unsuccessful metamorphosis of recently settled pediveligers into postlarvae and juveniles (Bayne, 1965; Talmage and Gobler, 2009, 2010), reduced shell calcification (Clements and Hunt, 2017; Green et al., 2013, 2009, 2004), growth impairment (Parker et al., 2010) or a combination of these factors (Waldbusser et al., 2010).

The average biomass per individual (DW, AW and AFDW) varied among size-classes, but not among sampling sites. Given the relatively small contribution of the soft tissues (AFDW) to the overall biomass (DW), the pattern observed for relative ash weight (%AW) seems to have been mainly driven by variations in the mineral fraction of the shells (i.e., calcium carbonate content). Negative net-calcification rates have frequently been observed for infaunal and epifaunal marine bivalves, in response to elevated CO₂ (see reviews by Gazeau et al., 2013 and Clements and Hunt, 2017). Although the relative contributions of accretion and dissolution to this net-effect cannot usually be disentangled, several authors suggest increased post-deposition dissolution as the main cause for the observed losses of shell mass (Findlay et al., 2011; Nienhuis et al., 2010; Range et al., 2012; Rodolfo-Metalpa et al., 2011; Tunnicliffe et al., 2009). In fact, accretion depends on both

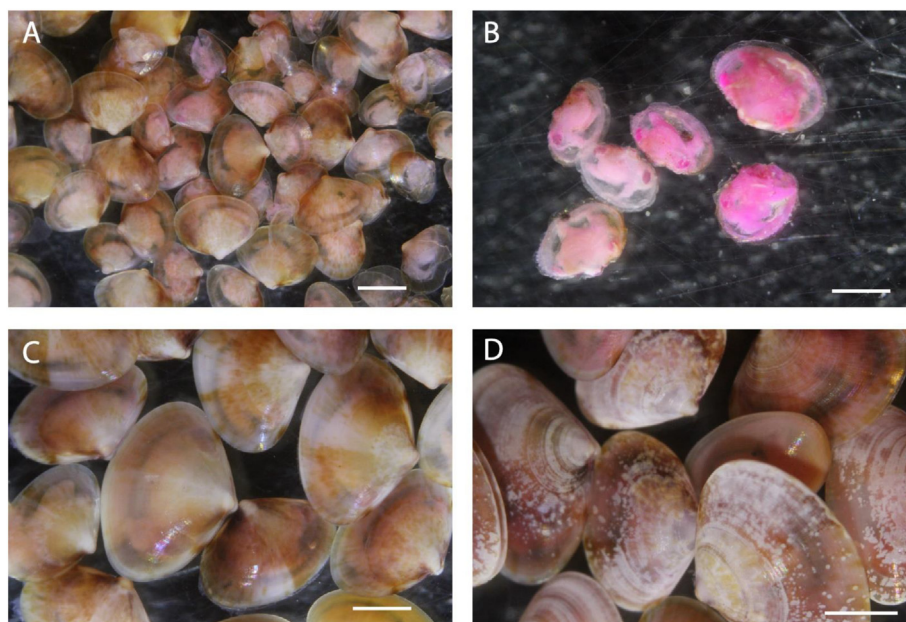


Fig. 5. Microphotographs of specimens of *Ervilia castanea* from different size classes and CO₂ levels: A (1–2 mm, reference), B (1–2 mm, high CO₂), C (2–3 mm, reference), D (2–3 mm, high CO₂). Scale bar is 1 mm.

Table 4

Sediment characteristics (means \pm SE) by island and location. Granulometry (<63 μ m, 63 μ m, 125 μ m, 250 μ m, 500 μ m, 1 mm, 2 mm and 4 mm), organic matter (OM), carbonate content (CaCO₃), total organic carbon (TOC) and total nitrogen (TN) are expressed as percentages. Other pigments include diadinoxanthin, diatoxanthin, zeaxanthin and lutein. C:N ratio was not independently determined, but calculated, so it was not considered in the DistLM models.

	São Miguel			Faial		
	Reference (n = 9)	Intermediate (n = 9)	High (n = 9)	Reference (n = 6)	Intermediate (n = 6)	High (n = 10)
<63 μ m	0.127 \pm 0.032	0.130 \pm 0.072	0.102 \pm 0.007	0.327 \pm 0.080	0.071 \pm 0.012	0.061 \pm 0.010
63 μ m	2.970 \pm 0.857	0.263 \pm 0.053	1.980 \pm 0.375	7.420 \pm 0.598	0.783 \pm 0.613	0.245 \pm 0.043
125 μ m	37.00 \pm 7.220	15.60 \pm 2.080	38.10 \pm 2.200	45.00 \pm 1.570	19.40 \pm 3.450	24.50 \pm 1.480
250 μ m	40.40 \pm 4.560	59.90 \pm 2.390	48.80 \pm 0.827	22.00 \pm 1.600	42.20 \pm 6.320	45.70 \pm 4.010
500 μ m	15.2 \pm 4.61	20.4 \pm 3.61	9.34 \pm 1.38	8.04 \pm 1.17	2.36 \pm 0.59	1.3 \pm 0.063
1 mm	2.8 \pm 1.09	3.29 \pm 0.818	1.35 \pm 0.268	7.02 \pm 0.983	6.44 \pm 0.869	8.78 \pm 1.26
2 mm	0.595 \pm 0.237	0.381 \pm 0.117	0.137 \pm 0.036	3.82 \pm 0.458	13 \pm 3.23	13.1 \pm 2.17
4 mm	0.438 \pm 0.347	0.039 \pm 0.031	0.006 \pm 0.006	5.91 \pm 2.82	15.6 \pm 4.93	6.34 \pm 2.11
OM (LOI ₅₅₀)	1.149 \pm 0.053	0.769 \pm 0.030	1.046 \pm 0.034	3.124 \pm 0.168	1.073 \pm 0.038	1.18 \pm 0.071
CaCO ₃ (LOI ₉₀₀)	0.477 \pm 0.043	0.219 \pm 0.035	0.574 \pm 0.042	6.516 \pm 0.393	0.085 \pm 0.018	0.209 \pm 0.048
Fucoxanthin (μ g/g)	0.426 \pm 0.051	0.095 \pm 0.013	0.063 \pm 0.014	0.211 \pm 0.023	0.04 \pm 0.007	0.054 \pm 0.027
Chl a (μ g/g)	1.097 \pm 0.13	0.231 \pm 0.029	0.169 \pm 0.047	0.717 \pm 0.116	0.072 \pm 0.019	0.12 \pm 0.064
Other pigments (μ g/g)	0.008 \pm 0.008	0	0	0.041 \pm 0.005	0	0
TN	0.077 \pm 0.015	0.067 \pm 0.012	0.06 \pm 0.011	0.061 \pm 0.013	0.076 \pm 0.012	0.079 \pm 0.010
TOC	0.181 \pm 0.014	0.142 \pm 0.010	0.166 \pm 0.029	0.256 \pm 0.032	0.159 \pm 0.018	0.138 \pm 0.006
C:N ratio	2.44 \pm 0.338	2.100 \pm 0.201	2.665 \pm 0.499	5.036 \pm 0.972	2.348 \pm 0.402	1.947 \pm 0.266

carbonate saturation of the surrounding water and the physiological status of the organism, which is largely determined by food availability, whereas dissolution relates solely to the external saturation state (Marshall et al., 2019).

Dissolution of the shell is recognized as an important source of post-settlement mortality in bivalves, having been associated with the

exponential loss of individuals after the transition from the larval to the benthic stage (Green et al., 2004). The type of shell damage observed at sites affected by CO₂ emissions has been previously reported in several laboratory (Bressan et al., 2014; Gazeau et al., 2014; Range et al., 2012) and in situ studies (Rodolfo-Metalpa et al., 2011; Thomsen et al., 2010). Other studies have also found that increased CO₂ levels are often associated with decreases in the mechanical properties of the shell, such as, thickness (Bressan et al., 2014; Gaylord et al., 2011; Talmage and Gobler, 2010), density (Klok et al., 2014; Rühl et al., 2017), hardness and fracture resistance (Beniash et al., 2010; Dickinson et al., 2013; Gazeau et al., 2013; Welladsen et al., 2010). Thinner or weaker shells have also been shown to increase bivalve mortality through several pathways, mediated by interactions with other environmental stressors (Dickinson et al., 2012), predators (Grey et al., 2007; Sanford et al., 2014), competitors (Coleman et al., 2014) or parasites (MacLeod and Poulin, 2015).

A number of recent studies have shown that the detrimental effects of elevated CO₂ on the calcification and growth of bivalves can be mitigated or avoided when the food supply is not limiting (Melnzer et al., 2011; Pansch et al., 2014; Ramajo et al., 2016b, 2016a; Thomsen et al., 2013). We did not assess if food limitation occurred during our study, but sediments at reference sites had greater amounts of chlorophyll *a* and inorganic carbon, relative to sites affected by CO₂ emissions (Fig. 6, Table 4). This implies that reference sites were better feeding grounds for infaunal bivalves. Concentrations of chlorophyll-*a* in seawater showed a similar pattern, with the smallest values consistently being found at the High CO₂ sites (unpublished data). Chlorophyll-*a* was, therefore, negatively correlated with CO₂ level and positively correlated with abundance and size of *E. castanea*. This strongly suggests that the indirect effects of CO₂, mediated by trophic interactions (i.e., food availability), may be equally or even more important than its direct effects on growth and survival of these clams.

Exposure to elevated CO₂ during development can also lead to reduced fitness and increased post-settlement mortality, due only to reductions in growth rates and energetic reserves (Hettinger et al., 2012; Waldbusser et al., 2015). Some of these mechanisms probably contributed to the observed effects on the population densities of *E. castanea*. Inferring on the relative importance of direct and indirect effects would, however, require manipulative experiments, which was clearly beyond the scope of the present study.

Hyp. 3 was also corroborated. In contrast to the overall pattern, abundances of the smaller size classes of *E. castanea* (<1 mm in São Miguel and <2 mm in Faial) generally increased with rising CO₂ levels. This pattern is clear, despite the abundance of spat (<1 mm) probably being underestimated, given that size at settlement is smaller than the

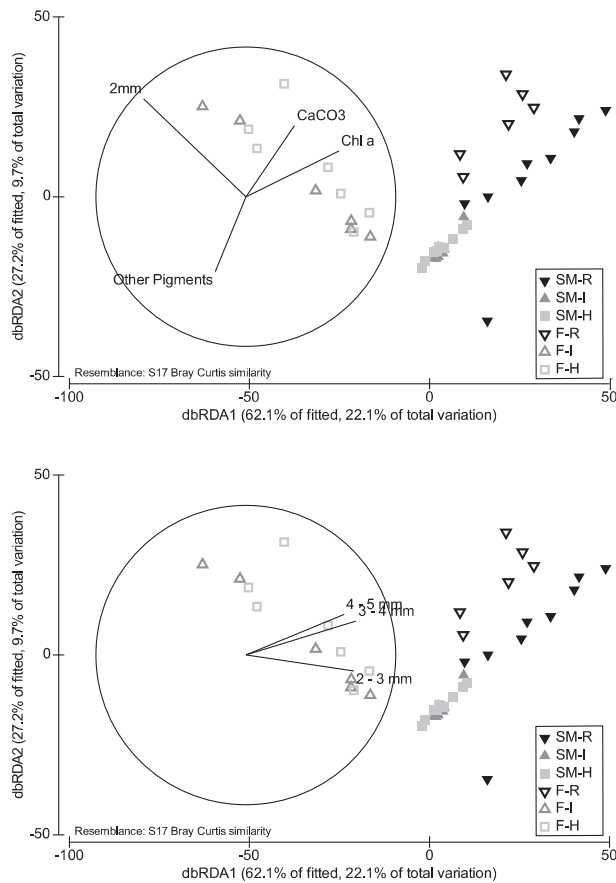


Fig. 6. Distance-based redundancy analysis (dbRDA) for the best solution of distance-based linear model (DistLM). Vector overlays represent significant variables included in the model, selected with the BEST procedure and Akaike's information criterion; top panel represents explanatory (sediment) variables and bottom panel the response variables (abundances of *E. castanea* per size class); the length of the vectors represents the relative influence of each variable to the ordination; São Miguel – SM, F – Faial, Reference, I – Intermediate CO₂, H – High CO₂.

500 µm mesh used for sampling (Bayne, 1965; Chicharo and Chicharo, 2001). Although larval settlement was not directly measured, this suggests that settlement rates were higher in sites affected by CO₂ emissions than in reference sites. The fact that newly settled postlarvae were found only in high CO₂ sites (Fig. 5B) also seems to corroborate this hypothesis.

Other populations of the same genus are known to have extreme temporal variations in abundance (i.e., boom-and-bust dynamics - Albano et al., 2016), which implies highly variable settlement rates. Potential spatial or temporal differences in settlement among experimental sites could, therefore, also have influenced the abundance and size-frequency distribution of the populations. The preponderance of small bivalves in naturally acidified environments has previously been observed and interpreted as evidence of continuous recruitment (Metaxas, 2015). As far as we are aware, however, there is no experimental evidence for positive effects of CO₂ on larval settlement of calcifying benthic marine invertebrates (Gazeau et al., 2013; Parker et al., 2013), while neutral (Bechmann et al., 2011; Crim et al., 2011) or negative effects (Cigliano et al., 2010; Ricevuto et al., 2012) have been observed. Accordingly, a direct enhancement of bivalve settlement by elevated levels of CO₂ seems extremely unlikely. Any positive effect was probably indirect, regulated by density-dependent control mechanisms.

Intraspecific interactions, like competition for resources (i.e., food or space) or predation (i.e., oophagy and larviphagy), have previously been recognized as important regulating mechanisms for population size in filter-feeding bivalves, by imposing a negative relationship between recruitment success and densities of adult conspecifics (Andre et al., 1993; Andre and Rosenberg, 1991; Comtet and Desbruyères, 1998; Vânia et al., 2014). It seems likely, therefore, that the high densities of adults in reference sites could lead to reduced rates of recruitment.

Chemical cues released by adult conspecifics are known to induce larval settlement and metamorphosis in several species of bivalves (Gosling, 2015; Porri et al., 2007). While the terms settlement and metamorphosis are often used interchangeably (Hadfield and Paul, 2001), they actually designate separate processes: metamorphosis is a definitive morphogenetic event; settlement, on the other hand, is reversible, as pediveligers and plantigrades of some species can swim up from the benthos and settle again in a new location (Bayne, 1964; Hadfield and Paul, 2001). Given the absence of *E. castanea* adults, chemical cues may be lacking from sites with elevated CO₂, which could have contributed to delayed metamorphosis and/or increased post-settlement dispersal, while having no effect on larval settlement itself. Although these potential mechanisms were not investigated in our study, the simultaneous occurrence of high levels of intraspecific predation (in reference sites) and the lack of chemical cues from adult conspecifics (in sites affected by CO₂) could explain the observed trends in the abundance of postlarvae and juveniles.

According to Morton (1990), *Ervilia castanea* in the Azores reach sexual maturity at a shell length of 3.5 mm for females and 5.5 mm for males. The survival threshold in sites with elevated levels of CO₂ was clearly under that size range, suggesting that individuals at those sites are not producing offspring. Being an open system, however, larvae may originate from non-acidified sites, and only after settlement become subjected to the effects of elevated CO₂ (Dupont and Pörtner, 2013; Kroeker et al., 2013b). Overall, and even though reproduction was not investigated in our study, these results suggest that populations of *E. castanea* in high CO₂ sites are probably sink populations.

5. Conclusions

One of the advantages of conducting ocean acidification research using natural CO₂ gradients is that the responses observed are not restricted to the direct effect of carbonate chemistry, but also include the indirect consequences of trophic interactions and changes in habitat complexity (Fabricius et al., 2015; Hale et al., 2011; Range et al., 2012). In the future, transplant experiments including measurements of

mineralization and dissolution of the shell, physiological, metabolic and genetic endpoints could enlighten and complement the responses observed in this study. There is also a need to consider other stressors (e.g., temperature, nutritional conditions, pollution), as future ecosystem changes will be driven by the synergistic effects of multiple stressors, instead of their isolated action (Baggini et al., 2014; Hale et al., 2011; Kroeker et al., 2013b). Future work should also include environmental benthic variables, such as sediment pH and temperature, as these can significantly vary from water column values (Clements et al., 2016; Dashfield et al., 2008).

We have shown that shallow-water volcanic CO₂ emissions in the Azores can be used to assess the effects of ocean acidification in central Atlantic benthic ecosystems, complementing similar studies in the region (Viotti et al., 2019) and in other parts of the world (Fabricius et al., 2015; Hall-Spencer et al., 2008; Harvey et al., 2018). Our results also emphasized the importance of food availability (i.e., chlorophyll *a*) in mediating the effects of CO₂ on filter feeders. Although *Ervilia castanea* is not commercially exploited, these findings raise concerns about the effects of ocean acidification on coastal bivalve populations with similar life history traits and on coastal economies that depend on them for food provision or commercial exploitation.

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CRediT authorship contribution statement

Marta Martins: Investigation, Formal analysis, Visualization, Writing - original draft. **Marina Carreiro-Silva:** Funding acquisition, Conceptualization, Methodology, Resources, Formal analysis, Writing - review & editing, Project administration. **Gustavo M. Martins:** Funding acquisition, Conceptualization, Methodology, Resources, Investigation, Formal analysis, Writing - review & editing, Project administration. **Joana Barcelos e Ramos:** Funding acquisition, Conceptualization, Methodology, Investigation, Resources, Formal analysis, Writing - review & editing, Project administration. **Fátima Viveiros:** Funding acquisition, Conceptualization, Methodology, Resources, Formal analysis, Writing - review & editing, Project administration. **Ruben P. Couto:** Funding acquisition, Conceptualization, Methodology, Investigation, Formal analysis, Resources, Writing - review & editing, Project administration. **Hugo Parra:** Investigation, Formal analysis. **João Monteiro:** Investigation, Formal analysis, Writing - review & editing. **Francesca Gallo:** Methodology, Investigation, Formal analysis, Writing - review & editing. **Catarina Silva:** Investigation, Formal analysis, Writing - review & editing. **Alexandra Teodósio:** Funding acquisition, Resources, Investigation, Writing - review & editing. **Katja Guilini:** Methodology, Investigation, Resources, Writing - review & editing. **Jason M. Hall-Spencer:** Funding acquisition, Conceptualization, Methodology, Investigation, Writing - review & editing, Project administration. **Francisco Leitão:** Funding acquisition, Methodology, Investigation, Formal analysis, Writing - review & editing, Supervision. **Luís Chicharo:** Funding acquisition, Resources, Writing - review & editing, Supervision. **Pedro Range:** Funding acquisition, Conceptualization, Methodology, Investigation, Formal analysis, Visualization, Writing - review & editing, Project administration, Supervision.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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References

- Agostini, S., Harvey, B.P., Wada, S., Kon, K., Milazzo, M., Inaba, K., Hall-Spencer, J.M., 2018. Ocean acidification drives community shifts towards simplified non-calcified habitats in a subtropical—temperate transition zone. *Sci. Rep.* 8, 1–11. <https://doi.org/10.1038/s41598-018-29251-7>.
- Albano, P.G., Filippova, N., Steger, J., Kaufman, D.S., Tomašových, A., Stachowitsch, M., Zuschin, M., 2016. Oil platforms in the Persian (Arabian) Gulf: living and death assemblages reveal no effects. *Cont. Shelf Res.* 121, 21–34. <https://doi.org/10.1016/j.csr.2015.12.007>.
- Amaral, V., Cabral, H.N., Bishop, M.J., 2011. Resistance among wild invertebrate populations to recurrent estuarine acidification. *Estuar. Coast. Shelf Sci.* 93, 460–467. <https://doi.org/10.1016/j.ejcs.2011.05.024>.
- Andre, C., Rosenberg, R., 1991. Adult-larval interactions in the suspension-feeding bivalves *Cerastoderma edule* and *Mya arenaria*. *Mar. Ecol. Prog. Ser.* 71, 227–234. <https://doi.org/10.3354/meps071227>.
- Andre, C., Jonsson, P.R., Lindgarth, M., 1993. Predation on settling bivalve larvae by benthic suspension feeders: the role of hydrodynamics and larval behaviour. *Mar. Ecol. Prog. Ser.* 97, 183–192. <https://doi.org/10.3354/meps097183>.
- Babio, C.R., Bonnin, J.F., 1987. Distribución y redescubrimiento de *Ervillea castanea* (Montagu, 1803) (Mollusca, Bivalvia, Eulamellibranchia, Mesodesmatidae) de las costas de Europa.
- Baggini, C., Salomidi, M., Voutsinas, E., Bray, L., Krasakopoulou, E., Hall-Spencer, J.M., 2014. Seasonality affects macroalgal community response to increases in pCO₂. *PLoS One* 9, e106520. <https://doi.org/10.1371/journal.pone.0106520>.
- Barros, P., Sobral, P., Range, P., Chicharo, L., Matias, D., 2013. Effects of sea-water acidification on fertilization and larval development of the oyster *Crassostrea gigas*. *J. Exp. Mar. Biol. Ecol.* 440, 200–206. <https://doi.org/10.1016/j.jembe.2012.12.014>.
- Bayne, B.L., 1964. Primary and secondary settlement in *Mytilus edulis* L. (Mollusca). *J. Anim. Ecol.* 33, 513–523. <https://doi.org/10.2307/2569>.
- Bayne, B.L., 1965. Growth and the delay of metamorphosis of the larvae of *Mytilus edulis* (L.). *Ophelia* 2, 1–47. <https://doi.org/10.1080/00785326.1965.10409596>.
- Beckmann, R.K., Taban, I.C., Westerlund, S., Godal, B.F., Arnberg, M., Vingen, S., Ingvarsdotter, A., Baussant, T., 2011. Effects of ocean acidification on early life stages of shrimp (*Pandalus borealis*) and mussel (*Mytilus edulis*). *Journal of Toxicology and Environmental Health - Part A: Current Issues*, 424–438. <https://doi.org/10.1080/15287394.2011.550460>.
- Beniash, E., Ivanina, A., Lieb, N., Kurochkin, I., Sokolova, I., 2010. Elevated level of carbon dioxide affects metabolism and shell formation in oysters *Crassostrea virginica* (Gmelin). *Mar. Ecol. Prog. Ser.* 419, 95–108. <https://doi.org/10.3354/meps08841>.
- Bindoff, N.L., Cheung, W.W.L., Kairo, J.G., Aristegui, J., Guinder, V.A., Hallberg, R., Hilmi, N., Jiao, N., Karim, M.S., Levin, L., O'Donoghue, S., Purca Cuicapusa, S.R., Rinkevich, B., Suga, T., Tagliabue, A., Williamson, P., 2019. In: Pörtner, H.-O., Roberts, D.C., Masson-Delmotte, V., Zhai, P., Tignor, M., Poloczanska, E., Mintenbeck, K., Alegría, A., Nicolai, M., Okem, A., Petzold, J., Rama, B., We, N.M. (Eds.), 2019: Changing Ocean, Marine Ecosystems, and Dependent Communities, IPCC Special Report: The Ocean and Cryosphere in a Changing Climate Summary for Policymakers <https://www.ipcc.ch/report/srocc/>.
- Boatta, F., D'Alessandro, W., Gagliano, A.L., Liotta, M., Milazzo, M., Rodolfo-Metalpa, R., Hall-Spencer, J.M., Parello, F., 2013. Geochemical survey of Levante Bay, Vulcano Island (Italy), a natural laboratory for the study of ocean acidification. *Mar. Pollut. Bull.* 73, 485–494. <https://doi.org/10.1016/j.marpolbul.2013.01.029>.
- Bressan, M., Chinellato, A., Munari, M., Matozzo, V., Mancini, A., Marčeta, T., Finos, L., Moro, I., Pastore, P., Badocco, D., Marin, M.G., 2014. Does seawater acidification affect survival, growth and shell integrity in bivalve juveniles? *Mar. Environ. Res.* 99, 136–148. <https://doi.org/10.1016/j.marenvres.2014.04.009>.
- Caliro, S., Viveiros, F., Chiodini, G., Ferreira, T., 2015. Gas geochemistry of hydrothermal fluids of the S. Miguel and Terceira Islands, Azores. *Geochim. Cosmochim. Acta* 168, 43–57.
- Calosi, P., Rastrick, S.P.S., Graziano, M., Thomas, S.C., Baggini, C., Carter, H.A., Hall-Spencer, J.M., Milazzo, M., Spicer, J.L., 2013. Distribution of sea urchins living near shallow water CO₂ vents is dependent upon species acid-base and ion-regulatory abilities. *Mar. Pollut. Bull.*, Ecological impacts of CCS leakage 73, 470–484. <https://doi.org/10.1016/j.marpolbul.2012.11.040>.
- Carriker, M.R., 1961. Interrelation of functional morphology, behavior and autecology in early stages of the bivalve *Mercenaria mercenaria*. *Journal of the Elisha Mitchell Scientific Society* 77, 168–241.
- Chicharo, L., Chicharo, M.A., 2001. Effects of environmental conditions on planktonic abundances, benthic recruitment and growth rates of the bivalve mollusc *Ruditapes decussatus* in a Portuguese coastal lagoon. *Fish. Res.* 53, 235–250. [https://doi.org/10.1016/S0165-7836\(00\)00290-3](https://doi.org/10.1016/S0165-7836(00)00290-3).
- Christen, N., Calosi, P., McNeill, C.L., Widdicombe, S., 2013. Structural and functional vulnerability to elevated pCO₂ in marine benthic communities. *Mar. Biol.* 160, 2113–2128. <https://doi.org/10.1007/s00227-012-2097-0>.
- Cigliano, M., Gambi, M.C., Rodolfo-Metalpa, R., Patti, F.P., Hall-Spencer, J.M., 2010. Effects of ocean acidification on invertebrate settlement at volcanic CO₂ vents. *Mar. Biol.* 157, 2489–2502. <https://doi.org/10.1007/s00227-010-1513-6>.
- Clements, J.C., Hunt, H.L., 2017. Effects of CO₂-driven sediment acidification on infaunal marine bivalves: a synthesis. *Mar. Pollut. Bull.* <https://doi.org/10.1016/j.marpolbul.2017.01.053> Elsevier Ltd.
- Clements, J.C., Woodard, K.D., Hunt, H.L., 2016. Porewater acidification alters the burrowing behavior and post-settlement dispersal of juvenile soft-shell clams (*Mya arenaria*). *J. Exp. Mar. Biol. Ecol.* 477, 103–111. <https://doi.org/10.1016/j.jembe.2016.01.013>.
- Clements, J.C., Carver, C.E., Mallet, M.A., Comeau, L.A., Mallet, A.L., 2020. CO₂-induced low pH in an eastern oyster (*Crassostrea virginica*) hatchery positively affects reproductive development and larval survival but negatively affects larval shape and size, with no intergenerational linkages. *ICES J. Mar. Sci.*, 0 <https://doi.org/10.1093/icesjms/fsaa089>.
- Coleman, D., Byrne, M., Davis, A., 2014. Molluscs on acid: gastropod shell repair and strength in acidifying oceans. *Mar. Ecol. Prog. Ser.* 509, 203–211. <https://doi.org/10.3354/meps10887>.
- Comtet, T., Desbruyères, D., 1998. Population structure and recruitment in mytilid bivalves from the Lucky Strike and Menez Gwen hydrothermal vent fields (37°17'N and 37°50'N on the Mid-Atlantic Ridge). *Mar. Ecol. Prog. Ser.* 163, 165–177. <https://doi.org/10.3354/meps163165>.
- Couto, R.P., Rodrigues, A.S., Neto, A.I., 2015. Shallow-water hydrothermal vents in the Azores (Portugal). *J. Integr. Coast. Zo. Manag.* 15, 495–505. <https://doi.org/10.5894/rgeci584>.
- Crim, R.N., Sunday, J.M., Harley, C.D.G., 2011. Elevated seawater CO₂ concentrations impair larval development and reduce larval survival in endangered northern abalone (*Haliotis kamtschatkana*). *J. Exp. Mar. Biol. Ecol.* 400, 272–277. <https://doi.org/10.1016/j.jembe.2011.02.002>.
- Cyronak, T., G. Schulz, K., L. Jokiel, P., 2016. The Omega myth: what really drives lower calcification rates in an acidifying ocean. *ICES J. Mar. Sci.* 73, 558–562. <https://doi.org/10.1093/icesjms/fsv075>.
- Dashfield, S.L., Somerfield, P.J., Widdicombe, S., Austen, M.C., Nimmo, M., 2008. Impacts of ocean acidification and burrowing urchins on within-sediment pH profiles and subtidal nematode communities. *J. Exp. Mar. Biol. Ecol.* 365, 46–52. <https://doi.org/10.1016/j.jembe.2008.07.039>.
- Dickinson, G.H., Ivanina, A.V., Matoo, O.B., Pörtner, H.O., Lannig, G., Bock, C., Beniash, E., Sokolova, I.M., 2012. Interactive effects of salinity and elevated CO₂ levels on juvenile eastern oysters, *Crassostrea virginica*. *J. Exp. Biol.* 215, 29–43. <https://doi.org/10.1242/jeb.061481>.
- Dickinson, G.H., Matoo, O.B., Tourek, R.T., Sokolova, I.M., Beniash, E., 2013. Environmental salinity modulates the effects of elevated CO₂ levels on juvenile hard-shell clams, *Mercenaria mercenaria*. *J. Exp. Biol.* 216, 2607–2618. <https://doi.org/10.1242/jeb.082909>.
- Dickson, A.G., 2010. Standards for ocean measurements. *Oceanog* 23, 34–47. <https://doi.org/10.5670/oceanog.2010.22>.
- Dickson, A.G., Millero, F.J., 1987. A comparison of the equilibrium constants for the dissociation of carbonic acid in seawater media. *Deep Sea Res. Part A: Oceanogr. Res. Pap.* 34, 1733–1743. [https://doi.org/10.1016/0198-0149\(87\)90021-5](https://doi.org/10.1016/0198-0149(87)90021-5).
- Dickson, A.G., Afghani, J.D., Anderson, G.C., 2003. Reference materials for oceanic CO₂ analysis: a method for the certification of total alkalinity. *Mar. Chem.* 80, 185–197. [https://doi.org/10.1016/S0304-4203\(02\)00133-0](https://doi.org/10.1016/S0304-4203(02)00133-0).
- Doney, S.C., Ruckelshaus, M., Emmett Duffy, J., Barry, J.P., Chan, F., English, C.A., Galindo, H.M., Grebmeier, J.M., Hollowed, A.B., Knowlton, N., Polovina, J., Rabalais, N.N., Sydeman, W.J., Talley, L.D., 2012. Climate change impacts on marine ecosystems. *Annu. Rev. Mar. Sci.* 4, 11–37. <https://doi.org/10.1146/annurev-marine-041911-111611>.
- Duarte, C.M., Hendriks, I.E., Moore, T.S., Olsen, Y.S., Steckbauer, A., Ramajo, L., Carstensen, J., Trotter, J.A., McCulloch, M., 2013. Is ocean acidification an open-ocean syndrome? Understanding anthropogenic impacts on seawater pH. *Estuar. Coasts*, 1–16 <https://doi.org/10.1007/s12237-013-9594-3>.
- Dupont, S., Pörtner, H., 2013. Marine science: get ready for ocean acidification. *Nature* 498, 429. <https://doi.org/10.1038/498429a>.
- Dupont, S., Thorndyke, M.C., 2009. Impact of CO₂-driven ocean acidification on invertebrates early life-history: what we know, what we need to know and what we can do. *Biogeosci. Discuss.* 6, 3109–3131.
- Dupont, S., Dorey, N., Stumpp, M., Melzner, F., Thorndyke, M., 2012. Long-term and trans-life-cycle effects of exposure to ocean acidification in the green sea urchin *Strongylocentrotus droebachiensis*. *Mar. Biol.*, 1–9 <https://doi.org/10.1007/s00227-012-1921-x>.
- Durland, E., Waldbusser, G., Langdon, C., 2019. Comparison of larval development in domesticated and naturalized stocks of the Pacific oyster *Crassostrea gigas* exposed to high pCO₂ conditions. *Mar. Ecol. Prog. Ser.* 621, 107–125. <https://doi.org/10.3354/meps12983>.
- Fabrizius, K.E., De'ath, G., Noonan, S., Uthicke, S., 2015. Ecological effects of ocean acidification and habitat complexity on reef-associated macroinvertebrate communities.

- Proc. R. Soc. London B Biol. Sci. 281, 20132479. <https://doi.org/10.1098/rspb.2013.2479>.
- Feely, R.A., Doney, S.C., Cooley, S.R., 2009. Ocean acidification: present conditions and future changes in a high-CO₂ world. *Oceanography. The Future of Ocean Biogeochemistry in a High-CO₂ World* 22, 26–35.
- Fernández-Reiriz, M.J., Range, P., Álvarez-Salgado, X.A., Labarta, U., 2011. Physiological energetics of juvenile clams *Ruditapes decussatus* in a high CO₂ coastal ocean. *Mar. Ecol. Prog. Ser.* 433, 97–105. <https://doi.org/10.3354/meps09062>.
- Fernández-Reiriz, M.J., Range, P., Álvarez-Salgado, X.A., Espinosa, J., Labarta, U., 2012. Tolerance of juvenile *Mytilus galloprovincialis* to experimental seawater acidification. *Mar. Ecol. Prog. Ser.* 454, 65–74. <https://doi.org/10.3354/meps09660>.
- Ferreira, T., Gaspar, J.L., Viveiros, F., Marcos, M., Faria, C., Sousa, F., 2005. Monitoring of fumarole discharge and CO₂ soil degassing in the Azores: contribution to volcanic surveillance and public health risk assessment. *Ann. Geophys.* 48, 787–796.
- Findlay, H.S., Wood, H.L., Kendall, M.A., Widdicombe, S., Spicer, J.I., Twitchett, R.J., 2011. Comparing the impact of high CO₂ on calcium carbonate structures in different marine organisms. *Mar. Biol. Res.* 7, 565–575. <https://doi.org/10.1080/17451000.2010.547200>.
- Garilli, V., Rodolfo-Metalpa, R., Scuderi, D., Brusca, L., Parrinello, D., Rastrick, S.P.S., Foggo, A., Twitchett, R.J., Hall-Spencer, J.M., Milazzo, M., 2015. Physiological advantages of dwarfing in surviving extinctions in high-CO₂ oceans. *Nat. Clim. Chang.*, 1–6. <https://doi.org/10.1038/nclimate2616>.
- Gaylord, B., Hill, T.M., Sanford, E., Lenz, E.A., Jacobs, L.A., Sato, K.N., Russell, A.D., Hettinger, A., 2011. Functional impacts of ocean acidification in an ecologically critical foundation species. *J. Exp. Biol.* 214, 2586–2594. <https://doi.org/10.1242/jeb.055939>.
- Gazeau, F., Gattuso, J.P., Dawber, C., Pronker, A.E., Peene, F., Peene, J., Heip, C.H.R., Middelburg, J.J., 2010. Effect of ocean acidification on the early life stages of the blue mussel *Mytilus edulis*. *Biogeosciences* 7, 2051–2060. <https://doi.org/10.5194/bg-7-2051-2010>.
- Gazeau, F., Parker, L.M., Comeau, S., Gattuso, J.-P., O'Connor, W.A., Martin, S., Pörtner, H.-O., Ross, P.M., O'Connor, W.A., Martin, S., Pörtner, H.-O., Ross, P.M., 2013. Impacts of ocean acidification on marine shelled molluscs. *Mar. Biol.* 160, 2207–2245. <https://doi.org/10.1007/s00227-013-2219-3>.
- Gazeau, F., Allouane, S., Bock, C., Bramanti, L., Correa, M.L., Gentile, M., Hirse, T., Pörtner, H.O., Ziveri, P., 2014. Impact of ocean acidification and warming on the Mediterranean mussel (*Mytilus galloprovincialis*). *Front. Mar. Sci.* 1. <https://doi.org/10.3389/fmars.2014.00062>.
- Gosling, E., 2015. *Marine Bivalve Molluscs: Second Edition*. Second Edition. Wiley Blackwell, Marine Bivalve Molluscs <https://doi.org/10.1002/9781119045212>.
- Green, M.A., Jones, M.E., Boudreau, C.L., Moore, R.L., Westman, B.A., 2004. Dissolution mortality of juvenile bivalves in coastal marine deposits. *Limnol. Oceanogr.* 49, 727–734. <https://doi.org/10.4319/lo.2004.49.3.0727>.
- Green, M.A., Waldbusser, G.G., Reilly, S.L., Emerson, K., O'Donnell, S., 2009. Death by dissolution: sediment saturation state as a mortality factor for juvenile bivalves. *Limnol. Oceanogr.* 54, 1037–1047. <https://doi.org/10.4319/lo.2009.54.4.1037>.
- Green, M.A., Waldbusser, G.G., Hubacz, L., Cathcart, E., Hall, J., 2013. Carbonate mineral saturation state as the recruitment cue for settling bivalves in marine muds. *Estuar. Coasts* 36, 18–27. <https://doi.org/10.1007/s12237-012-9549-0>.
- Grey, M., Lelievre, P.G., Boulding, E.G., 2007. Selection for prey shell thickness by the naticid gastropod *Euspira lewisii* (Naticidae) on the bivalve *Protothaca staminea* (Veneridae). *Veliger* 48, 317–322.
- Hadfield, M., Paul, V., 2001. Natural chemical cues for settlement and metamorphosis of marine-invertebrate larvae. In: McClintock, J., Baker, B. (Eds.), *Marine Chemical Ecology*, Marine Science. CRC Press, pp. 431–461. <https://doi.org/10.1201/9781420036602.ch13>.
- Hale, R., Calosi, P., McNeill, L., Mieszkowska, N., Widdicombe, S., 2011. Predicted levels of future ocean acidification and temperature rise could alter community structure and biodiversity in marine benthic communities. *Oikos* 120, 661–674. <https://doi.org/10.1111/j.1600-0706.2010.19469.x>.
- Hall-Spencer, J.M., Harvey, B.P., 2019. Ocean acidification impacts on coastal ecosystem services due to habitat degradation. *Emerging Topics in the Life Sciences* 3, 197–206.
- Hall-Spencer, J.M., Rodolfo-Metalpa, R., Martin, S., Ransome, E., Fine, M., Turner, S.M., Rowley, S.J., Tedesco, D., Buia, M.-C., Jason, M., Hall-Spencer, R.R.-M., 2008. Volcanic carbon dioxide vents show ecosystem effects of ocean acidification. *Nature* 454, 96–99. <https://doi.org/10.1038/nature07051>.
- Hansen, H.P., Koroleff, F., 1999. Determination of nutrients. In: Grasshoff, K., Kremling, K., Ehrhardt, M. (Eds.), *Methods of Seawater Analysis*, Third edition Wiley-VCH Verlag GmbH, Weinheim, Germany <https://doi.org/10.1002/9783527613984.ch10>.
- Harvey, B.P., Agostini, S., Wada, S., Inaba, K., Hall-Spencer, J.M., 2018. Dissolution: the Achilles' heel of the triton shell in an acidifying ocean. *Front. Mar. Sci.* 5. <https://doi.org/10.3389/fmars.2018.00371>.
- Heiri, O., Lotter, A.F., Lemcke, G., 2001. Loss on ignition as a method for estimating organic and carbonate content in sediments: reproducibility and comparability of results. *J. Paleolimnol.* 25, 101–110. <https://doi.org/10.1023/A:1008119611481>.
- Hettinger, A., Sanford, E., Hill, T.M., Russell, A.D., Sato, K.N.S., Hoey, J., Forsch, M., Page, H.N., Gaylord, B., 2012. Persistent carry-over effects of planktonic exposure to ocean acidification in the Olympia oyster. *Ecology* 93, 2758–2768. <https://doi.org/10.1890/12-0567.1>.
- Hoegh-Guldberg, O., Cai, R., Poloczanska, E.S., Brewer, P.G., Sundby, S., Hilmi, K., Fabry, V.J., Jung, S., 2014. The ocean. In: Barros, V.R., Field, C.B., Dokken, D.J., Mastrandrea, M.D., Mach, K.J., Bilir, T.E., Chatterjee, M., Ebi, K.L., Estrada, Y.O., Genova, R.C., Girma, B., Kissel, E.S., Levy, A.N., MacCracken, S., Mastrandrea, P.R., White, L.L. (Eds.), *Climate Change 2014: I*. Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA, pp. 1655–1731.
- Jansson, A., Lischka, S., Boxhammer, T., Schulz, K.G., Norkko, J., 2015. Larval development and settling of *Macoma balthica* in a large-scale mesocosm experiment at different fCO₂ levels. *Biogeosci. Discuss.* 12, 20411–20435. <https://doi.org/10.5194/bgd-12-20411-2015>.
- Keeling, Ralph, F., Keeling, C.D., 2017. Atmospheric monthly in situ CO₂ data - Mauna Loa Observatory, Hawaii. [WWW document]. Atmos. Mon. Situ CO₂ Data - Mauna Loa Obs. Hawaii. Scripps CO₂ Progr. Data. UC San Diego Libr. Digit. Collect <https://doi.org/10.6075/J08W3BHW>.
- Klok, C., Wijsman, J.W.M., Kaag, K., Foekema, E., 2014. Effects of CO₂ enrichment on cockle shell growth interpreted with a Dynamic Energy Budget model. *J. Sea Res.* 94, 111–116. <https://doi.org/10.1016/j.seares.2014.01.011>.
- Kroeker, K.J., Kordas, R.L., Crim, R.N., Singh, G.G., 2010. Meta-analysis reveals negative yet variable effects of ocean acidification on marine organisms. *Ecol. Lett.* 13, 1419–1434. <https://doi.org/10.1111/j.1461-0248.2010.01518.x>.
- Kroeker, K.J., Micheli, F., Gambi, M.C., Martz, T.R., 2011. Divergent ecosystem responses within a benthic marine community to ocean acidification. *Proc. Natl. Acad. Sci. U. S. A.* 108, 14515–14520. <https://doi.org/10.1073/pnas.1107789108>.
- Kroeker, K.J., Gambi, M.C., Micheli, F., Cristina, M., Micheli, F., 2013a. Community dynamics and ecosystem simplification in a high-CO₂ ocean. *Proc. Natl. Acad. Sci. U. S. A.* 110, 12721–12726. <https://doi.org/10.1073/pnas.1216464110>.
- Kroeker, K.J., Kordas, R.L., Crim, R., Hendriks, I.E., Ramajo, L., Singh, G.S., Duarte, C.M., Gattuso, J.-P., 2013b. Impacts of ocean acidification on marine organisms: quantifying sensitivities and interaction with warming. *Glob. Chang. Biol.* 19, 1884–1896. <https://doi.org/10.1111/gcb.12179>.
- Lewis, E.R., Wallace, D.W.R., 1998. Program developed for CO₂ system calculations (no. cdiac:CDIAC-105). Environmental System Science Data Infrastructure for a Virtual Ecosystem <https://doi.org/10.15485/1464255>.
- Macedo, M.C.C., Macedo, M.I.C., Borges, J.P., 1999. *Conchas Marinhas de Portugal = Sea-shells of Portugal*. Verbo, Lisboa.
- MacLeod, C.D., Poulin, R., 2015. Differential tolerances to ocean acidification by parasites that share the same host. *Int. J. Parasitol.* 45, 485–493. <https://doi.org/10.1016/j.ijpara.2015.02.007>.
- Marčeta, T., Matozzo, V., Alban, S., Badocco, D., Pastore, P., Marin, M.G., 2020. Do males and females respond differently to ocean acidification? An experimental study with the sea urchin *Paracentrotus lividus*. *Environ. Sci. Pollut. Res.* <https://doi.org/10.1007/s11356-020-10040-7>.
- Marshall, D.J., Abdelhady, A.A., Wah, D.T.T., Mustapha, N., Gödeke, S.H., De Silva, L.C., Hall-Spencer, J.M., 2019. Biomonitoring acidification using marine gastropods. *Sci. Total Environ.* 692, 833–843. <https://doi.org/10.1016/j.scitotenv.2019.07.041>.
- Mehrbach, C., Culberson, C.H., Hawley, J.E., Pytkowicz, R.M., 1973. Measurement of the apparent dissociation constants of carbonic acid in seawater at atmospheric pressure. *Limnol. Oceanogr.* 18, 897–907. <https://doi.org/10.4319/lo.1973.18.6.0897>.
- Mellado, C., Chaparro, O.R., Duarte, C., Villanueva, P.A., Ortiz, A., Valdivia, N., Torres, R., Navarro, J.M., 2019. Ocean acidification exacerbates the effects of paralytic shellfish toxins on the fitness of the edible mussel *Mytilus chilensis*. *Sci. Total Environ.* 653, 455–464. <https://doi.org/10.1016/j.scitotenv.2018.10.399>.
- Melzner, F., Stange, P., Trübenbach, K., Thomsen, J., Casties, I., Panknin, U., Gorb, S.N., Gutowska, M.A., 2011. Food supply and seawater pCO₂ impact calcification and internal shell dissolution in the blue mussel *Mytilus edulis*. *PLoS One* 6, e24223. <https://doi.org/10.1371/journal.pone.0024223>.
- Metaxas, A., 2015. Bivalve populations inhabiting hydrothermal vents on submarine volcanoes: using size frequency distributions to infer potential regulatory factors. *Mar. Ecol.* 36, 62–70. <https://doi.org/10.1111/maec.12248>.
- Mishra, A.K., Santos, R., Hall-Spencer, J.M., 2020. Elevated trace elements in sediments and seagrass at CO₂ seeps. *Mar. Environ. Res.* 153, 104810.
- Morton, B., 1990. The biology and functional morphology of *Erilia castanea* (Bivalvia: Tellinacea) from the Azores. *Açoreana* 75–96.
- Morton, B., Da Cunha, R.T., De Frias Martins, A.M., 2013. Species richness, relative abundance and dwarfism in Azorean bivalves: consequences of latitude, isolation or productivity? Or all three? *J. Mar. Biol. Assoc. United Kingdom* 94, 1–12. <https://doi.org/10.1017/S0025315413001598>.
- Nienhuis, S., Palmer, A.R., Harley, C.D.G., 2010. Elevated CO₂ affects shell dissolution rate but not calcification rate in a marine snail. *Proc. R. Soc. B Biol. Sci.* 277, 2553–2558. <https://doi.org/10.1098/rspb.2010.0206>.
- Palmer, A.R., 1992. Calcification in marine molluscs: how costly is it? *Proc. Natl. Acad. Sci. U. S. A.* 89, 1379–1382. <https://doi.org/10.1073/pnas.89.4.1379>.
- Pansch, C., Schaub, I., Havenhand, J., Wahl, M., 2014. Habitat traits and food availability determine the response of marine invertebrates to ocean acidification. *Glob. Chang. Biol.* 20, 765–777. <https://doi.org/10.1111/gcb.12478>.
- Parker, L.M., Ross, P.M., O'Connor, W.A., 2010. Comparing the effect of elevated pCO₂ and temperature on the fertilization and early development of two species of oysters. *Mar. Biol.* 157, 2435–2452. <https://doi.org/10.1007/s00227-010-1508-3>.
- Parker, L.M., Ross, P.M., O'Connor, W.A., Borysko, L., Raftos, D.A., Pörtner, H., 2012. Adult exposure influences offspring response to ocean acidification in oysters. *Glob. Chang. Biol.* 18, 82–92. <https://doi.org/10.1111/j.1365-2486.2011.02520.x>.
- Parker, L., Ross, P., O'Connor, W., Pörtner, H., Scanes, E., Wright, J., 2013. Predicting the response of molluscs to the impact of ocean acidification. *Biology (Basel)* 2, 651–692. <https://doi.org/10.3390/biology2020651>.
- Pereira, A.M., Range, P., Campoy, A., Oliveira, A.P., Joaquim, S., Matias, D., Chicharro, L., Gaspar, M.B., 2015. Larval hatching and development of the wedge shell (*Donax trunculus* L.) under increased CO₂ in southern Portugal. *Reg. Environ. Chang.* 16, 855–864. <https://doi.org/10.1007/s10113-015-0803-4>.
- Porri, F., Zardi, G.I., McQuaid, C.D., Radloff, S., 2007. Tidal height, rather than habitat selection for conspecifics, controls settlement in mussels. *Mar. Biol.* 152, 631–637. <https://doi.org/10.1007/s00227-007-0716-y>.
- Pörtner, H., 2008. Ecosystem effects of ocean acidification in times of ocean warming: a physiologist's view. *Mar. Ecol. Prog. Ser.* 373, 203–217. <https://doi.org/10.3354/meps07768>.

- Pörtner, H.-O., Karl, D., Boyd, P.W., Cheung, W., Lluch-Cota, S.E., Njorri, Y., Schmidt, D.N., Zavalov, P., 2014. Ocean systems. In: Field, C.B., Barros, V.R., Dokken, D.J., Mach, K.J., Mastrandrea, M.D., Bilir, T.E., Chatterjee, M., Ebi, K.L., Estrada, Y.O., Genova, R.C., Girma, B., Kissel, E.S., Levy, A.N., MacCracken, S., Mastrandrea, P.R., White, L.L. (Eds.), *Climate Change 2014: I*. Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA, pp. 411–484.
- Rajasabapathy, R., Mohandass, C., Colaco, A., Dastager, S.G., Santos, R.S., Meena, R.M., 2014. Culturable bacterial phylogeny from a shallow water hydrothermal vent of Espalamaca (Faial, Azores) reveals a variety of novel taxa. *Curr. Sci.* 106, 58–69.
- Ramajo, L., Marbà, N., Prado, L., Peron, S., Lardies, M.A., Rodriguez-Navarro, A.B., Vargas, C.A., Lagos, N.A., Duarte, C.M., 2016a. Biomineralization changes with food supply confer juvenile scallops (*Argopecten purpuratus*) resistance to ocean acidification. *Glob. Chang. Biol.* 22, 2025–2037. <https://doi.org/10.1111/gcb.13179>.
- Ramajo, L., Pérez-León, E., Hendriks, I.E., Marbà, N., Krause-Jensen, D., Sejr, M.K., Blicher, M.E., Lagos, N.A., Olsen, Y.S., Duarte, C.M., 2016b. Food supply confers calcifiers resistance to ocean acidification. *Sci. Rep.* 6, 1–6. <https://doi.org/10.1038/srep19374>.
- Range, P., Pilo, D., Ben-Hamadou, R., Chicharo, M.A., Matias, D., Joaquim, S., Oliveira, A.P.P., Chicharo, L., Piló, D., Ben-Hamadou, R., Chicharo, M.A.A., Matias, D., Joaquim, S., Oliveira, A.P.P., Chicharo, L., 2012. Seawater acidification by CO₂ in a coastal lagoon environment: effects on life history traits of juvenile mussels *Mytilus galloprovincialis*. *J. Exp. Mar. Bio. Ecol.* 424–425, 89–98. <https://doi.org/10.1016/j.jembe.2012.05.010>.
- Range, P., Chicharo, M.A., Ben-Hamadou, R., Piló, D., Fernandez-Reiriz, M.J., Labarta, U., Marin, M.G., Bressan, M., Matozzo, V., Chinellato, A., Munari, M., Menif, N.T.E., Dellali, M., Chicharo, L., 2014a. Impacts of CO₂-induced seawater acidification on coastal Mediterranean bivalves and interactions with other climatic stressors. *Reg. Environ. Chang.* 14, 19–30. <https://doi.org/10.1007/s10113-013-0478-7>.
- Range, P., Martins, M., Cabral, S., Piló, D., Ben-Hamadou, R., Teodósio, M., Leitão, F., Drago, T., Oliveira, A., Matias, D., Chicharo, L., 2014b. Relative sensitivity of soft-bottom intertidal macrofauna to increased CO₂ and experimental stress. *Mar. Ecol. Prog. Ser.* 509, 153–170. <https://doi.org/10.3354/meps10861>.
- Ricevuto, E., Lorenti, M., Patti, F.P., Scipione, M.B., Gambi, M.C., 2012. Temporal trends of benthic invertebrate settlement along a gradient of ocean acidification at natural CO₂ vents (Tyrrhenian sea). *Biol. Mar. Mediterr.* 19, 49–52.
- Rodolfo-Metalpa, R., Houlbrèque, É., Boisson, F., Baggini, C., Patti, F.P., Jeffree, R., Fine, M., Foggo, A., Gattuso, J.-P.P., Hall-Spencer, J.M., 2011. Coral and mollusc resistance to ocean acidification adversely affected by warming. *Nat. Clim. Chang.* 1, 308–312. <https://doi.org/10.1038/nclimate1200>.
- Roleda, M.Y., Boyd, P.W., Hurd, C.L., 2012. Before ocean acidification: calcifier chemistry lessons. *J. Phycol.* 48, 840–843. <https://doi.org/10.1111/j.1529-8817.2012.01195.x>.
- Rühl, S., Calosi, P., Faulwetter, S., Keklikoglou, K., Widdicombe, S., Queirós, A.M., 2017. Long-term exposure to elevated pCO₂ more than warming modifies early-life shell growth in a temperate gastropod. *ICES J. Mar. Sci.* 74, 1113–1124. <https://doi.org/10.1093/icesjms/fsw242>.
- Sanford, E., Gaylord, B., Hettinger, A., Lenz, E.A., Meyer, K., Hill, T.M., 2014. Ocean acidification increases the vulnerability of native oysters to predation by invasive snails. *Proc. R. Soc. B Biol. Sci.* 281, 20132681. <https://doi.org/10.1098/rspb.2013.2681>.
- Shirayama, Y., 2005. Effect of increased atmospheric CO₂ on shallow water marine benthos. *J. Geophys. Res.* 110. <https://doi.org/10.1029/2004JC002618>.
- Talmage, S.C., Gobler, C.J., 2009. The effects of elevated carbon dioxide concentrations on the metamorphosis, size, and survival of larval hard clams (*Mercenaria mercenaria*), bay scallops (*Argopecten irradians*), and Eastern oysters (*Crassostrea virginica*). *Limnol. Oceanogr.* 54, 2072–2080.
- Talmage, S.C., Gobler, C.J., 2010. Effects of past, present, and future ocean carbon dioxide concentrations on the growth and survival of larval shellfish. *Proc. Natl. Acad. Sci.* 107, 17246–17251. <https://doi.org/10.1073/pnas.0913804107>.
- Tarasov, V.G., Gebruk, A.V., Mironov, A.N., Moskalov, L.I., 2005. Deep-sea and shallow-water hydrothermal vent communities: two different phenomena? *Chemical Geology, Shallow-Water Hydrothermal Venting* 224, 5–39. <https://doi.org/10.1016/j.chemgeo.2005.07.021>.
- Thomsen, J., Gutowska, M.A., Saphörster, J., Heinemann, a., Trübenbach, K., Fietzke, J., Hiebenthal, C., Eisenhauer, a., Körtzinger, a., Wahl, M., Melzner, F., 2010. Calcifying invertebrates succeed in a naturally CO₂-rich coastal habitat but are threatened by high levels of future acidification. *Biogeosciences* 7, 3879–3891. <https://doi.org/10.5194/bg-7-3879-2010>.
- Thomsen, J., Casties, I., Pansch, C., Körtzinger, A., Melzner, F., 2013. Food availability outweighs ocean acidification effects in juvenile *Mytilus edulis*: laboratory and field experiments. *Glob. Chang. Biol.* 19, 1017–1027. <https://doi.org/10.1111/gcb.12109>.
- Tribollet, A., Grange, J.S., Parra, H., Rodolfo-Metalpa, R., Carreiro-Silva, M., 2018. Limited carbonate dissolution by boring microflora at two volcanically acidified temperate sites: Ischia (Italy, Mediterranean Sea) and Faial (Azores, NE Atlantic Ocean). *Glob. Biogeochem. Cycles* 32, 78–91. <https://doi.org/10.1002/2016GB005575>.
- Tunncliffe, V., Davies, K.T.A., Butterfield, D.A., Embley, R.W., Rose, J.M., Chadwick, W.W., 2009. Survival of mussels in extremely acidic waters on a submarine volcano. *Nat. Geosci.* 2, 344–348. <https://doi.org/10.1038/ngeo500>.
- Vânia, B., Ullah, H., Teixeira, C.M., Range, P., Erzini, K., Leitão, F., 2014. Influence of environmental variables and fishing pressure on bivalve fisheries in an inshore lagoon and adjacent nearshore coastal area. *Estuar. Coasts* 37, 191–205. <https://doi.org/10.1007/s12237-013-9658-4>.
- Vihtakari, M., Hendriks, I., Holding, J., Renaud, P., Duarte, C., Havenhand, J., 2013. Effects of ocean acidification and warming on sperm activity and early life stages of the Mediterranean mussel (*Mytilus galloprovincialis*). *Water* 5, 1890–1915. <https://doi.org/10.3390/w5041890>.
- Viotti, S., Sangil, C., Hernández, C.A., Hernández, J.C., 2019. Effects of long-term exposure to reduced pH conditions on the shell and survival of an intertidal gastropod. *Mar. Environ. Res.* 152, 104789. <https://doi.org/10.1016/j.marenvres.2019.104789>.
- Viveiros, F., Moreno, L., Carreiro-Silva, M., Couto, R., Silva, C., Range, P., Galdes, D., Parra, H., Ferreira, T., Pacheco, J., 2016. Volcanic gas emissions offshore of São Miguel and Faial islands (Azores archipelago). *Actas das 4as Jornadas Eng. Hidrográica* 320–323.
- Vizzini, S., Di Leonardo, R., Costa, V., Tramati, C.D., Luzzu, F., Mazzola, A., 2013. Trace element bias in the use of CO₂ vents as analogues for low pH environments: implications for contamination levels in acidified oceans. *Estuar. Coast. Shelf Sci.* 134, 19–30. <https://doi.org/10.1016/j.ecss.2013.09.015>.
- Waldbusser, G.G., Bergschneider, H., Green, M.A., 2010. Size-dependent pH effect on calcification in post-larval hard clam *Mercenaria* spp. *Mar. Ecol. Prog. Ser.* 417, 171–182. <https://doi.org/10.3354/meps08809>.
- Waldbusser, G.G., Hales, B., Langdon, C.J., Haley, B.A., Schrader, P., Brunner, E.L., Gray, M.W., Miller, C.A., Gimenez, I., Hutchinson, G., 2015. Ocean acidification has multiple modes of action on bivalve larvae. *PLoS One* 10. <https://doi.org/10.1371/journal.pone.0128376>.
- Wallenstein, F.M., Couto, R.P., Amaral, A.S., Wilkinson, M., Neto, A.I., Rodrigues, A.S., 2009. Baseline metal concentrations in marine algae from São Miguel (Azores) under different ecological conditions - urban proximity and shallow water hydrothermal activity. *Mar. Pollut. Bull.* 58, 438–443. <https://doi.org/10.1016/j.marpolbul.2008.11.021>.
- Wallenstein, F.M., Couto, R.P., Torrão, D.F., Neto, A.I., Rodrigues, A.S., Wilkinson, M., 2013. Intertidal rocky shore seaweed communities subject to the influence of shallow water hydrothermal activity in São Miguel (Azores, Portugal). *Helgol. Mar. Res.* 67, 535–543. <https://doi.org/10.1007/s10152-012-0341-0>.
- Welladsen, H.M., Southgate, P.C., Heimann, K., 2010. The effects of exposure to near-future levels of ocean acidification on shell characteristics of *Pinctada fucata* (Bivalvia: Pteriidae). *Molluscan Res.* 30, 125–130.
- Widdicombe, S., Spicer, J.I., 2008. Predicting the impact of ocean acidification on benthic biodiversity: what can animal physiology tell us? *J. Exp. Mar. Bio. Ecol.* 366, 187–197. <https://doi.org/10.1016/j.jembe.2008.07.024>.
- Widdicombe, S., Dashfield, S.L., McNeill, C.L., Needham, H.R., Beesley, a., McEvoy, a., Øxnevad, S., Clarke, K.R., Berge, Ja., 2009. Effects of CO₂ induced seawater acidification on infaunal diversity and sediment nutrient fluxes. *Mar. Ecol. Prog. Ser.* 379, 59–75. <https://doi.org/10.3354/meps07894>.
- Wright, S.W., Jeffrey, S.W., 1997. High resolution HPLC system for chlorophylls and carotenoids of marine phytoplankton. In: Jeffrey, S.W., Mantoura, R.F.C., Wright, S.W. (Eds.), *Phytoplankton Pigments in Oceanography*. UNESCO Publishing, Paris, pp. 327–341.
- Zhao, L., Yang, F., Milano, S., Han, T., Walliser, E.O., Schöne, B.R., 2018. Transgenerational acclimation to seawater acidification in the Manila clam *Ruditapes philippinarum*: preferential uptake of metabolic carbon. *Sci. Total Environ.* 627, 95–103. <https://doi.org/10.1016/j.scitotenv.2018.01.225>.
- Zhao, L., Liu, B., An, W., Deng, Y., Lu, Y., Liu, B., Wang, L., Cong, Y., Sun, X., 2019. Assessing the impact of elevated pCO₂ within and across generations in a highly invasive fouling mussel (*Musculista senhousia*). *Sci. Total Environ.* 689, 322–331. <https://doi.org/10.1016/j.scitotenv.2019.06.466>.